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(54) Title: LYMPHOTOXIN-β, LYMPHOTOXIN-β COMPLEXES, PHARMACEUTICAL PREPARATIONS AND THERAPEUTIC USES THEREOF			
(57) Abstract			
<p>This invention relates to lymphotoxin-β, a lymphocyte membrane type protein. This protein is found on the surface of a number of cells, including phorbol ester (PMA) stimulated T cell hybridoma II-23.D7 cells. This invention also relates to complexes formed between lymphotoxin-β and other peptides such as lymphotoxin-α and to complexes comprising multiple subunits of lymphotoxin-β. These proteins and complexes are useful in holding LT-α formed within the cell on the cell surface where the LT-α/LT-β complex may act as an inflammation regulating agent, a tumor growth inhibiting agent, a T cell inhibiting agent, a T cell activating agent, an autoimmune disease regulating agent, or an HIV inhibiting agent. Furthermore, the antitumor activity of the LT-α/LT-β complex may be delivered to tumor cells by tumor infiltrating lymphocytes (TILs) transfected with the gene for LT-β.</p>			

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**LYMPHOTOXIN- β , LYMPHOTOXIN- β COMPLEXES,
PHARMACEUTICAL PREPARATIONS AND
THERAPEUTIC USES THEREOF**

TECHNICAL FIELD OF THE INVENTION

5 This invention relates to lymphotoxin- β , a lymphocyte membrane-type polypeptide. Lymphotoxin- β , also referred to as p33, has been identified on the surface of T lymphocytes, T cell lines, B cell lines and lymphokine-activated killer cells.

10 This invention also relates to complexes formed between lymphotoxin- β (LT- β) and other lymphotoxin-type peptides such as lymphotoxin (which we refer to herein as "lymphotoxin- α (LT- α)" to distinguish it from LT- β) and to complexes comprising
15 multiple subunits of LT- β . The LT- β polypeptide of this invention is expected to be useful in holding LT- α formed within the cell on the cell surface where either LT- β or the LT- α /LT- β complex may act as an inflammation regulating agent, a tumor growth
20 inhibiting agent, a T cell inhibiting agent, a T cell activating agent, an immunomodulatory agent, an autoimmune disease regulating agent or an HIV regulating agent. Furthermore, the antitumor activity of the LT- α /LT- β complex may be delivered to tumor
25 cells by tumor infiltrating lymphocytes (TILs) transfected with the gene for LT- β .

 The invention described herein was made in part during the course of work under Grant

SUBSTITUTE SHEET (RULE 26)

- 2 -

No. CA 35638-07-10 from the National Institutes of Health. The U.S. Government has certain rights in this invention.

BACKGROUND OF THE INVENTION

5 The initiation of the immune response involves a complex array of intercellular signals. These signals typically involve soluble cytokines coupled with a number of cell-cell contact dependent signals. The contact dependent events, most notably
10 activation of the T-cell receptor, lend specificity to the response whereas the soluble mediators are generally responsible for maintenance of cell differentiation and proliferation. Tumor Necrosis Factor (TNF) and LT- α are two polypeptides generally
15 recognized for involvement with the initiation of the immune response.

 TNF and LT- α are soluble proteins noted originally for their ability to inhibit the growth of tumors. [L. Old, "Tumor Necrosis Factor," Science,
20 230, 630 (1985)]. Further research demonstrated that both proteins exhibit a wide range of activities. TNF is synthesized in response to various inflammatory insults by a variety of cell types including both hematopoietic and nonhematopoietic cells, while LT- α ,
25 in contrast, is made specifically by lymphocytes. The two known TNF receptors do not appear to discriminate between LT- α and TNF. [T. Schall et al., "Molecular Cloning and Expression of a Receptor for Human Tumor Necrosis Factor," Cell, 61, 361-370 (1990); C. Smith
30 et al., "A Receptor for Tumor Necrosis Factor Defines an Unusual Family of Cellular and Viral Proteins," Science 248, 1019 (1990)]. In general, LT- α and TNF display similar spectra of activities in in vitro systems, although LT- α is often less potent.

SUBSTITUTE SHEET (RULE 26)

- 3 -

[J. Browning et al., Studies Of The Differing Affects Of Tumor Necrosis Factor And Lymphotoxin On The Growth Of Several Human Tumor Lines," J. Immunol., 143, 1859 (1989)].

- 5 TNF appears to play a major role in specific aspects of metabolic control, in the response to endotoxin shock, and in the regulation of hematopoietic cell development. [B. Beutler et al., "The History, Properties, and Biological Effects of Cachectin,"
- 10 Biochemistry, 27, (1988); M. Akashi et al., "Lymphotoxin: Stimulation And Regulation of Colony Stimulating Factors in Fibroblasts," Blood, 74, 2383 (1989); G. Roodman et al., "Tumor Necrosis Factor-alpha and Hematopoietic Progenitors: Effects Of Tumor
- 15 Necrosis Factor On The Growth Of Erythroid Progenitors CFU-E And BFU-E And The Hematopoietic Cell Lines k562, HL60, And HEL Cells," Exp. Hematol., 15, 928 (1987)].

- Along with IL-1 and IL-6, TNF is also a major mediator of the inflammatory response. [D. Cavender
- 20 et al., "Endothelial Cell Activation Induced By Tumor Necrosis Factor And Lymphotoxin," Amer. Jour. Path., 134, 551 (1989); R. Cotran et al., "Endothelial Activation Its Role In Inflammatory And Immune Reactions," in Endothelial Cell Biology, (Plenum Press,
- 25 Simonescu & Simonescu, eds., 1988) 335]. TNF also appears to be involved in T cell activation under certain conditions. [M. Shalaby et al., "The Involvement Of Human Tumor Necrosis Factors- α And- β In The Mixed Lymphocyte Reaction," J. Immunol., 141, 499
- 30 (1988); N. Damle et al., "Distinct Regulatory Effects of IL-4 and TNF- α During CD3-Dependent and CD3-Independent Initiation Of Human T-Cell Activation," Lymph. Res., 8, 85 (1989); G. Ranges et al., "Tumor Necrosis Factor- α As A Proliferative Signal F r An IL-
- 35 2-Dependent T Cell Line: Strict Specif s Specificity of

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- 4 -

Action," Amer. Assoc. Immunol., 142, 1203 (1989);
G. Ranges et al., "Tumor Necrosis Factor α /Cachectin Is
A Growth Factor For Thymocytes," J. Exp. Med., 167,
1472 (1988); P. Scheurich et al., "Immunoregulatory
5 Activity Of Recombinant Human Tumor Necrosis Factor
(TNF)- α : Induction Of TNF Receptors On Human T Cells
And TNF- α -Mediated Enhancement Of T Cell Responses,"
J. Immunol., 138, 1786 (1987)].

TNF is produced by several types of cells,
10 including monocytes, fibroblasts, T cells and Natural
Killer (NK) cells. [D. Goeddel et al., "Tumor Necrosis
Factors: Gene Structure And Biological Activities,"
Cold Spring Harbor Symposium Quant. Biol., 51, 597
(1986); D. Spriggs et al., "Tumor Necrosis Factor
15 Expression In Human Epithelial Tumor Cell Lines,"
J. Clin. Invest., 81, 455 (1988); M. Turner et al.,
"Human T cells From Autoimmune and Normal Individuals
Can Produce Tumor Necrosis Factor," Eur. J. Immunol.,
17, 1807 (1987)]. Investigators have also detected
20 murine and human forms of TNF that are associated with
the surface of various cells either as a transmembrane
protein or a receptor-bound molecule. [B. Luettig
et al., "Evidence For the Existence of Two Forms of
Membrane Tumor Necrosis Factor: An Integral Protein
25 and a Molecule Attached To Its Receptor," J. Immunol.,
143, 4034 (1989); M. Kriegler et al., "A Novel Form of
TNF/Cachectin Is a Cell Surface Cytotoxic Transmembrane
Protein: Ramifications For the Complex Physiology of
TNF," Cell, 53, pp. 45-53 (1988); and M. Kinkhabwala
30 et al., "A Novel Addition To the T Cell Repertory,"
J. Exp. Med., 171, pp. 941-946 (1990)].

LT- α also has many activities, generally
similar, but not identical to those of TNF, including
tumor necrosis, induction of an antiviral state,
35 activation of polymorphonuclear leukocytes, induction

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- 5 -

- of class I major histocompatibility complex antigens on endothelial cells, induction of adhesion molecules on endothelium and growth hormone stimulation. [N. Ruddie and R. Homer, "The Role of Lymphotoxin in
- 5 Inflammation," Prog. Allergy, 40, pp. 162-182 (1988)]. Both LT- α and TNF are ligands to members of the nerve growth factor (NGF) receptor family. [S. Mallett and A.N. Barclay, "A New Superfamily Of Cell Surface
- 10 Immunology Today, 12, 7, 220-223 (1991).]

- In contrast to TNF, LT- α secretion appears to be a specific property of only activated T cells and certain B-lymphoblastoid tumors. [N. Paul et al., "Lymphotoxin," Ann. Rev. Immunol., 6, 407 (1988)].
- 15 Some researchers have also indicated that a membrane-associated form of LT- α may be expressed on the surface of lymphocytes under certain circumstances [J. Hiserodt, et al., "Identification of Membrane-Associated Lymphotoxin (LT) On Mitogen-Activated Human
- 20 Lymphocytes Using Heterologous Anti-LT Antisera In Vitro," Cell. Immunol., 34, pp. 326-339 (1977); C. Ware et al., "Mechanisms of Lymphocyte-Mediated Cytotoxicity," J. Immunol., 126, pp. 1927-1933 (1981); U. Anderson et al. J. Immunol. Methods, 123, 233
- 25 (1989); Y. Abe et al., Jpn. J. Canc. Res., 82, 23 (1991); Y. Abe et al., "Studies of Membrane Associated and Soluble (Secreted) Lymphotoxin In Human Lymphokine-Activated T-Killer Cells In Vitro," Lymphokine and Cytokine Research, 11, 2, 115-121 (1992)].

- 30 In recent years genes for both TNF and LT- α have been isolated and cloned, leading to their complete characterization and to the availability of recombinant forms of both proteins. [P. Gray et al., "Cloning and Expression of cDNA For Human Lymphotoxin,
- 35 A Lymphokine With Tumor Necrosis Activity," Nature,

SUBSTITUTE SHEET (RULE 26)

- 6 -

312, pp. 121-124 (1984); D. Pennica et al., "Human Tumor Necrosis Factor: Precursor Structure, Expression And Homology To Lymphotoxin," Nature, 312, 724 (1984)].

- Other "cytokine-like" cell surface proteins
- 5 including the CD40 protein have recently been shown to share certain similarities with TNF and LT- α . Like TNF and LT- α , the CD40 protein is a ligand to members of the TNF/nerve growth factor (NGF) receptor family. [S. Mallett and N. Barclay, Immunology Today, 12,
- 10 pp. 220-223 (1991)]. The CD40 protein is a 277-amino acid protein expressed on the surface of B lymphocytes, epithelial cells, and some carcinoma cell lines. [R. Armitage et al., Nature, 357, pp. 80-82 (1992); T. Farrah and C. Smith, "Emerging Cytokine Family,"
- 15 Nature, 358, p. 26 (1992)].

- We have now identified a novel surface protein, lymphotoxin- β (LT- β) or p33. LT- β has been identified on the surface of several types of lymphocyte cells, including OKT3-stimulated primary T
- 20 cells, antigen specific IL-2 dependent CTL clones, and a PMA-stimulated human T cell hybridoma II-23.D7. LT- β targets LT- α produced in the cell to the cell membrane where LT- β and LT- α appear as a complex (designated "LT- α /LT- β " throughout this disclosure). The LT- α /LT- β
- 25 complex is believed to be a novel mechanism for membrane expression of LT- α by activated T-cells. [Androlewicz et al., "Lymphotoxin Is Expressed As a Heteromeric complex With A Distinct 33 kDa Glycoprotein On The Surface Of An Activated Human T Cell Hybridoma,"
- 30 Journal Of Biological Chemistry, 267, pp. 2542-2547 (1992)]. The LT- α /LT- β complex may exhibit cytolytic and cell regulatory activity similar to the soluble LT- α , TNF and CD40 proteins. The membrane-associated LT- β complexed with LT- α may represent, as a complex, a

SUBSTITUTE SHEET (RULE 26)

- 7 -

novel ligand for T cell interactions with other cells and may also be useful in targeted cell lysis.

SUMMARY OF THE INVENTION

The novel protein of the present invention
5 has been named lymphotoxin- β (LT- β). This protein is found on the surface of several types of lymphocyte cells, including OKT3-stimulated primary T cells, antigen-specific IL-2 dependent CTL clones, and a PMA-stimulated human T cell hybridoma, II-23.D7. It forms
10 a novel complex with LT- α and forms complexes with other LT- β subunits (e.g., (LT- β)₂ LT- α complexes).

LT- β has a molecular weight of 31-35 kD as determined by immunoprecipitation and SDS-PAGE. LT- β exhibits N-linked glycosylation. The amino acid
15 sequence of lymphotoxin- β is set forth in SEQ ID NO:2, and the amino acid sequences of several soluble lymphotoxin- β peptides are set forth in SEQ ID NO:4, SEQ ID NO:6 and SEQ ID NO:8. The DNA sequence coding for lymphotoxin- β is set forth in SEQ ID NO:1 and DNA
20 sequences coding for several soluble lymphotoxin- β peptides are set forth in SEQ ID NO:3, SEQ ID NO:5, and SEQ ID NO:7.

LT- β as a cell membrane protein binds LT- α during synthesis, thus "targeting" the LT- α to the cell
25 membrane. In the absence of LT- β , LT- α is secreted into the extracellular medium. The LT- α /LT- β complex is recognized by polyclonal antisera raised against recombinant lymphotoxin- α (rLT- α) expressed in CHO cells or by monoclonal antibodies (mAbs) raised against
30 natural LT- α . Furthermore, antisera that recognize the LT- α /LT- β complex and (LT- α)₃ block the mixed lymphocyte reaction (MLR), a standard immunological assay of the expected proliferative response of T lymphocytes to allogenic stimulation, i.e., the

SUBSTITUTE SHEET (RULE 26)

- 8 -

introduction of T lymphocytes from another individual, which are recognized as foreign (non-self) by the "responder" lymphocytes. [See, e.g., M. Shalaby et al., J. Immunol., 141, 499 (1988)].

5 The LT- β protein was purified by affinity chromatography, partially sequenced and a specific oligonucleotide probe was designed. The cDNA encoding the LT- β was isolated by probing a cDNA library from activated II-23.D7 cells, a human T-cell hybridoma that
10 displays large amounts of surface lymphotoxin upon phorbol ester activation. The identified amino acid sequence encodes a 240-244 amino acid sequence (a molecular mass of the unmodified protein of about 25130-25390 kDa). See SEQ ID NO:2. The amino acid
15 sequence and the placement of the transmembrane region are typical of a type II membrane protein.

 This sequence comprises a short 14-18 amino acid N-terminal "cytoplasmic" domain. Following this cytoplasmic domain there is an extensive stretch of 30
20 hydrophobic amino acids which presumably acts as a membrane anchoring domain. No identical sequences were found within available databases. There is one cysteine residue in the extracellular domain and two methionines within the last C-terminal 17 amino acids.
25 This is consistent with the very limited cyanogen bromide cleavage pattern exhibited by this protein.

 Comparison of the LT- β sequence with other proteins known to bind to members of the TNF/NGF receptor family reveals considerable structural
30 similarity. Four of the ligands to members of the TNF/NGF receptor family (TNF, LT- α , LT- β and the CD40 ligand) resemble type II membrane proteins and share at least four regions of sequence conservation in the extracellular domain as indicated in Figure 14. The
35 conserved TNF and LT- α domains shared with LT- β are

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- 9 -

likely to be involved in intersubunit interactions and β sheet organization. These regions of conservation can account for the association between LT- α and LT- β . The existence of these homology regions may facilitate engineering the polypeptide to form complexes with, for example, TNF or the CD40 ligand. Such a molecule would have mixed functions and could possibly be used as a custom designed drug. [See J. Fuh et al., "Rational Design Of Potent Antagonists To The Human Growth Hormone Receptor," Science, 256, 1677 (1992)].

We believe that the polypeptide complexes of this invention are important in T cell activation events and are useful in compositions and methods for T cell activation or T cell suppression and as therapeutic agents in the treatment of inflammation or applications requiring cytolytic activities, such as inhibition of tumor cell or neoplasia growth. We also believe that the polypeptide complexes may be important in cellular immunotherapies, including enhancing the tumoricidal properties of tumor infiltrating lymphocytes in Tumor Infiltrating Lymphocyte ("TIL") therapy. TIL immunotherapy may be improved by gene transfer techniques. For example, a gene may be added to tumor cells for the purpose of inducing the body's immune system to mediate an effective tumor-directed immune response. [See, e.g., W.F. Anderson, "Human Gene Therapy." Science, 256, 808-813 (1992)].

We also believe, based upon similarities between a molecule identified as "Fas" and members of the TNF/NGF receptor family, that the polypeptide complexes of this invention may be involved in the internal cell process known as programmed cell death or apoptosis, and may therefore be involved in mediating autoimmune disease. [See, e.g., N. Itoh et al., "The Polypeptide Encoded By The cDNA For Human Cell Surface

SUBSTITUTE SHEET (RULE 26)

- 10 -

Antigen Fas Can Mediate Apoptosis," Cell, 66, 233-243
(1991); R. Watanabe-Fukunaga et al.,
"Lymphoproliferation Disorder In Mice Explained By
Defects In Fas Antigen That Mediates Apoptosis," Nature
5 356, 314 (1992)).

Antibodies to LT- β , its related polypeptides,
the LT- α /LT- β complex or the other polypeptide
complexes of this invention may also disrupt critical
LT interactions with particular receptors, thus
10 specifically affecting LT-mediated events other than
those mediated through the known TNF receptor forms.
Likewise, receptors for TNF, LT- α or LT- α /LT- β , or
their derivatives (e.g., soluble receptors and
IgG/receptor fusion proteins) may be used to inhibit
15 the polypeptides and complexes of this invention.

BRIEF DESCRIPTION OF THE DRAWINGS

Figure 1 depicts flow cytofluorometric
analysis of OKT3-stimulated, IL-2 expanded peripheral
blood lymphocytes (PBL) showing reaction with 3
20 different rabbit anti-rLT- α antisera and showing
essentially no reaction with rabbit anti-rTNF antisera.

Figure 2 depicts flow cytofluorometric
analysis of a human T cell hybridoma, II-23.D7, showing
the presence (following PMA treatment) of a LT- α -
25 related epitope on the cell surface.

Figure 3 depicts the ability of PMA activated
II-23.D7 cells to bind anti-rLT- α antibodies. Samples
of anti-rLT- α antisera were incubated with PMA-treated
U937 (non-LT-producing) cells, (-O-), PMA-activated II-
30 23.D7 hybridoma cells (10^8 , - - ; 10^7 , ---), and no
cells (control, - -). Serial dilutions of the cell-
free antisera after incubation were added to rLT- α and
used in a cytotoxicity assay against L929 (LT- α -
sensitive) cells. The plots indicate that rLT- α

SUBSTITUTE SHEET (RULE 26)

- 11 -

neutralizing antibodies were removed from the antisera by the activated II-23.D7 cells.

Figure 4 shows two autoradiographs (A and B) depicting immunoprecipitation of ^{125}I -labeled surface proteins from PMA-activated II-23.D7 cells. Figure 4A shows immunoprecipitation of an approximately 25 kD surface protein (LT- α) and an approximately 33 kD surface protein (p33, or LT- β) by post-immune rabbit anti-rLT- α antiserum but not by pre-immune rabbit serum. Figure 4B shows a 1-dimensional CNBr cleavage map of the 25 kD and 33 kD bands from figure 4A, compared against recombinant TNF (rTNF) produced in *E. coli*, and recombinant lymphotoxin- α (rLT- α) produced in CHO cells, [J. Browning et al., *J. Immunol.*, 143, 1859 (1989)] both with (+) and without (-) CNBr cleavage.

Figure 5 presents autoradiographs showing immunoprecipitation of TNF- and LT- α -related proteins from PMA-stimulated II-23.D7 cells metabolically labeled with ^{35}S -methionine or ^{35}S -cysteine. The figure shows recognition by anti-rLT- α antisera (L), but not preimmune (P) or anti-rTNF antisera (T), of an approximately 25 kD methionine-containing surface protein (LT- α) and an approximately 33 kD methionine- and cysteine-containing surface protein (p33, or LT- β). The autoradiographs also indicate that activated II-23.D7 cells also produce a 26 kD form of TNF and secrete soluble lymphotoxin- α .

Figure 6 depicts affinity purification 1-D CNBr peptide mapping of those proteins from PMA-activated II-23.D7 cells recognized by anti-rLT- α serum. Figure 6A represents SDS PAGE analysis of the proteins eluted from an anti-LT- α affinity column prepared from either pre-immune (PRE) or post-immune (POST) rabbit sera. Figure 6A shows the ~33kD and

SUBSTITUTE SHEET (RULE 26)

- 12 -

-20kD protein bands did not bind to an affinity column prepared using preimmune serum (PRE) but did bind to an affinity column prepared using anti-rLT- α antiserum (POST). Figure 6B shows partial CNBr cleavage of the
5 -33 kD and -20 kD proteins eluted from the POST column, compared against rTNF and rLT- α run in parallel. The gels were visualized by silver staining.

Figure 7 presents autoradiographs of the -25 kD and -33 kD 125 I-labeled proteins (designated LT- α
10 and LT- β , respectively) immunoprecipitated from activated II-23.D7 cells, treated with N-glycanase (N-gly), with a mixture of neuraminidase and O-glycanase (O-gly), or with all three enzymes.

Figure 8 depicts the results of a
15 reimmunoprecipitation of the coprecipitated p33 (LT- β) and p25 (LT- α) proteins to further investigate whether they are immunogenically related.

Figure 9 (comprising parts 9A and 9B) shows the results of isoelectric focusing under denaturing
20 conditions of the immunoprecipitated p33 (LT- β) and p25 (LT- α) proteins.

Figure 10 (comprising parts 10A and 10B) shows the results of isoelectric focusing under native conditions of the immunoprecipitated p33 (LT- β) and p25
25 (LT- α) proteins. Together Figs. 9 and 10 indicate that LT- β and LT- α form a denaturable complex.

Figure 11 depicts flow cytofluorometric analysis of surface proteins differentially expressed on T cells and monocytes after stimulation with a
30 mixture of LPS, IFN- γ and OKT3. From a stimulated PBL pool, separated T cells were observed to express a surface protein recognized by anti-rLT- α antisera (LT), whereas separated monocytes expressed a surface protein recognized by anti-rTNF antisera (TNF).

- 13 -

Figure 12 shows flow cytofluorometric analysis of surface LT forms on leu-19⁻ and leu-19⁺ (i.e., natural killer) cells treated with IL-2. Analysis of IL-2 treated PBL with both labeled leu-19
5 and anti-rLT- α confirms that lymphokine-activated killer (LAK) cells express a surface LT form.

Figure 13 depicts amino acid sequence of LT- β fragments obtained by direct N-terminal sequencing and in situ trypsin digestion followed by reverse phase
10 HPLC resolution of the digested peptides.

Figure 14 depicts an amino acid sequence comparison of four members of the family of ligands binding to members of the TNF/NGF-receptor family. Homology regions are shown in bold type face with
15 sequence identity indicated with a dot and conserved sequences with an asterisk. Putative N-linked glycosylation sites are boxed.

Figure 15 depicts northern analysis of LT- α and LT- β expression. A) Northern blot of several cell
20 lines showing specific expression of both LT genes in only Hut-78 and II-23.D7 cells. B) Time course of PMA induction of LT- α and LT- β mRNAs in II-23.D7 cells. C) Similar analysis of human peripheral blood lymphocytes activated with either anti-CD3 or IL-2
25 alone.

Figure 16 depicts expression of LT- α and LT- β in CHO cells. A) FACS analysis of CHO cells transiently transfected with the LT- β cDNA. Either parental dhFR-CHO or LT- α expressing CHO cells were
30 electroporated with either pCDM8 containing an irrelevant insert (clone 4) or the pCDM8/LT- β plasmid and stained with control IgG(---) or anti LT- α monoclonal antibodies (___). B) Panel B depicts expression of LT- α and LT- β in COS cells. Cells were
35 transfected with control DNA or the pCDM8/LT β plasmid,

SUBSTITUTE SHEET (RULE 26)

- 14 -

either with or without pCDM8/LTα and stained as per panel A.

DETAILED DESCRIPTION OF THE INVENTION

In order that the invention herein described
5 may be fully understood, the following detailed description is set forth.

This invention relates to lymphotoxin-β, a lymphocyte membrane-type polypeptide. The amino acid sequence of lymphotoxin-β is set forth in SEQ ID NO:2.

10 This polypeptide, also referred to as p33, has a molecular weight of 31 to 35 kD. The polypeptides of this invention may be associated with a cell surface or not associated with such a surface.

This invention also relates to soluble forms
15 of lymphotoxin-β. Soluble lymphotoxin-β peptides are defined by the amino acid sequence of lymphotoxin-β wherein the sequence is cleaved at any point between the end of the transmembrane region (i.e., at about amino acid #44) and the first homology region at about
20 amino acid #95. Amino acid sequences of two soluble lymphotoxin-β peptides are defined by SEQ ID NO:4 and SEQ ID NO:6. Several additional soluble lymphotoxin-β polypeptides comprise the amino acid sequence as defined by SEQ ID NO:6 with additional amino acid
25 residues at the 5' end. The additional residues may comprise the 52 amino acid residues as defined by SEQ ID NO:8. The soluble lymphotoxin may also comprise an amino acid sequence defined by SEQ ID NO:6 plus a portion of SEQ ID NO:8 comprising 1 to 51 of the amino
30 acid residues beginning from the 3' end. Such soluble peptides may include any number of well known leader sequences at the 5' end. Such a sequence would allow the peptides to be expressed in a eukaryotic system.

SUBSTITUTE SHEET (RULE 26)

- 15 -

[See, e.g., Ernst et al. U.S. Patent No. 5,082,783 (1992)].

The polypeptide complexes of this invention comprise a first polypeptide comprising an amino acid
5 sequence selected from the group of SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:6, and the sequence defined by SEQ ID NO:6 plus the entirety or a portion of SEQ ID NO:8 as described above, complexed with a second polypeptide
10 polypeptide such as: lymphotoxin- α , native human or animal lymphotoxin- α , recombinant lymphotoxin- α , soluble lymphotoxin- α , secreted lymphotoxin- α , or lymphotoxin- α -active fragments of any of the above.

The novel LT- β peptide forms a complex with
15 LT- α and forms complexes with other LT- β subunits (e.g., (LT- β)₂ LT- α complexes). These complexes may be cell associated or not associated with a cell, and may complex with other type II membrane proteins sharing common homology regions as described above.

20 The polypeptide complexes are recognized by polyspecific antisera that recognize recombinant LT- α , suggesting that the complex exhibits LT- α epitopes. These antisera include a commercial anti-LT- α monoclonal antibody (Boehringer Mannheim), and
25 polyspecific antisera raised against recombinant lymphotoxin- α (rLT- α) expressed in transfected Chinese Hamster Ovary (CHO) cells. Polyclonal antisera that recognize these complexes also block the mixed lymphocyte reaction (MLR), but a monoclonal anti-rLT- α
30 antibody that recognizes soluble LT- α do not block the MLR. The complexes of the present invention thus appear to play an important role in T cell activation. We also expect these complexes to have T cell
35 regulatory activities and cytotoxic activities similar to those of soluble LT- α or TNF.

SUBSTITUTE SHEET (RULE 26)

- 16 -

This invention also relates to DNA consisting essentially of DNA sequences that code for the polypeptides comprising the amino acid sequences described above, recombinant DNA molecules

5 characterized by that DNA, hosts selected from the group of unicellular hosts or animal and human cells in culture transfected with that DNA, and recombinant methods of using that DNA and those recombinant DNA molecules and hosts to produce the polypeptides coded

10 thereby.

More specifically, this invention relates to an isolated DNA sequence comprising the nucleotide sequence as defined by SEQ ID NO:1.

This invention also relates to polypeptides

15 encoded by that sequence, DNA sequences that hybridize with that DNA sequence that code for a polypeptide that is substantially homologous with lymphotoxin- β , and degenerate DNA sequences comprising nucleotide sequences that code for lymphotoxin- β .

20 This invention also relates to DNA sequences that code for soluble lymphotoxin- β peptides. These DNA sequences are defined by SEQ ID NO:3 and SEQ ID NO:5. This invention also relates to several additional soluble lymphotoxin- β peptides that are

25 coded for by the DNA of SEQ ID NO:5 plus several additional nucleotide triplets at the 5' end. The additional nucleotide triplets may comprise the 52 triplets as defined by SEQ ID NO:7. The soluble lymphotoxin peptide may also be encoded by SEQ ID NO:5

30 plus a portion of SEQ ID NO:7 comprising 1 to 51 nucleotide triplets beginning from the 3' end.

This invention also relates to DNA sequences that hybridize to any of the sequences identified above that code on expression for lymphotoxin- β or soluble

35 lymphot xin- β peptides. This invention also relates to

SUBSTITUTE SHEET (RULE 26)

- 17 -

degenerate nucleotide sequences that code for lymphotoxin- β or a soluble lymphotoxin- β peptide, and to DNA sequences that code for polypeptides that are substantially homologous with a soluble lymphotoxin- β peptide.

Lymphotoxin- β was identified, isolated and characterized using the techniques described below:

Flow Cytofluorometric Analysis

First we demonstrated the expression of LT- α epitopes on the surface of T cells using flow cytofluorometric analysis. We observed that human peripheral blood mononuclear cells activated with OKT3 monoclonal antibody demonstrated expression of LT- α epitopes by reacting with anti-rLT- α antisera. Only anti-rLT- α antisera, not anti-rTNF antisera, bound to OKT3-stimulated primary T cells.

We also observed that a human T cell hybridoma, II-23.D7 [C. Ware et al., "Human T Cell Hybridomas Producing Cytotoxic Lymphokines: Induction of Lymphotoxin Release And Killer Cell Activity By Anti-CD3 Monoclonal Antibody Or Lectins And Phorbol Ester," Lymph. Res., 5, 313 (1986)], secreted LT- α upon PMA stimulation and also expressed surface LT- α -related epitopes upon PMA stimulation. We also demonstrated that PMA-activated II-23.D7 cells were able to remove LT-neutralizing antibodies from the rabbit anti-rLT- α antisera, while control U937 cells, which lack all surface LT- α forms, were not. We further ruled out the possibility that the rabbit anti-rLT- α antisera had bound (complexed) with rabbit LT- α (the resulting complexes subsequently binding to cellular LT- α /TNF receptors on the II-23.D7 cells) by saturating the cellular receptors with excess soluble TNF or LT- α and observing that this had no effect on the staining.

SUBSTITUTE SHEET (RULE 26)

- 18 -

These assays demonstrate that the LT- α -related epitopes on this hybridoma are genuinely related to LT- α .

We also observed that pretreatment of the antisera with excess rLT- α blocked the ability of the
5 antisera to stain II-23.D7 cells, while pretreatment with rTNF had no effect. This assay demonstrated the specificity of the antisera for LT- α -related epitopes.

Trypsinization of the stimulated II-23.D7 cells prior to staining led to loss of the signal,
10 demonstrating that the epitopes recognized by the antisera were proteins.

We also demonstrated that CHO-derived contaminants did not contribute to the antisera recognition of induced proteins on the surface of
15 activated II-23.D7 cells by showing that CHO cells stably transfected with the LT- α gene, which produce only soluble LT- α , were not stained by the anti-LT- α antisera.

Immunoprecipitation

20 We further characterized these surface LT- α -related proteins by either surface iodination (125 I-labelling) or metabolic labelling (35 S-Met or 35 S-Cys) of PMA-activated II-23.D7 cells, followed by solubilization of the plasma membrane with detergent
25 and immunoprecipitation of the labeled LT- α -related proteins.

Surface iodination coupled with immunoprecipitation revealed two proteins recognized by the anti-LT- α antisera: a 25-26 kD form subsequently
30 referred to as LT- α , and a 31-35 kD form subsequently referred to as LT- β or p33. We observed that neither the preimmune serum from the same rabbit nor anti-rTNF rabbit serum immunoprecipitated these bands from the iodinated, PMA-activated II-23.D7 cells. One

SUBSTITUTE SHEET (RULE 26)

- 19 -

dimensional partial CNBr peptide mapping of the iodinated, immunoprecipitated bands showed that the 25-26 kD form (LT- α) cleaves in a pattern identical to that of iodinated recombinant LT- α , reinforcing the correlation between surface LT- α and soluble LT- α . The iodinated 31-35 kD form (LT- β , or p33) was not cleaved by CNBr, indicating that it is distinct from the known LT- α gene product.

We further characterized the LT- α and LT- β proteins by metabolic labelling of PMA-activated II-23.D7 cells with ^{35}S -methionine or ^{35}S -cysteine, followed by immunoprecipitation. The distribution of cysteine and methionine provides a means of distinguishing between TNF and LT- α and between forms of each with and without their signal sequences [M. Kriegler et al., Cell, 53, 45 (1988)]. Secreted TNF contains cysteine, but not methionine, while secreted LT- α contains only methionine and no cysteine residues. LT- α , however, has one cysteine residue in its signal sequence, while TNF contains two methionine residues in its signal sequence.

We labeled separate cultures of PMA-treated II-23.D7 cells with either ^{35}S -methionine or ^{35}S -cysteine and precipitated immunoreactive proteins from the culture media and the cells. Subsequent SDS-PAGE analysis of the immunoprecipitates from the culture media of cells labeled with ^{35}S -methionine revealed a 25 kD form of LT- α while the immunoprecipitates from the culture media of cells labeled with ^{35}S -cysteine did not, a pattern expected for secreted LT- α . Analysis of the washed cells showed both the 25-26 kD LT- α form and the 33 kD LT- β form. These results paralleled the membrane-associated forms observed using surface iodination.

- 20 -

The 25-26 kD LT- α lacked cysteine, indicating processing of the leader sequence. We also observed that the 33 kD LT- β incorporated both ^{35}S -methionine and ^{35}S -cysteine, distinguishing itself as different
5 from the 25 kD LT- α form. Typically, LT- α bound to its receptor can be cross-linked to the receptor using a chemical linker such as BOSCOES (i.e., (bis [2-[succinimidooxy-carboxyloxy]ethyl] sulfone; Pierce, Rockford, IL). [J.S. Andrews et al., "Characterization
10 of the Receptor for Tumor-Necrosis Factor (TNF) and Lymphotoxin (LT) on Human T Lymphocytes," J. Immunol., 144, 2582 (1990)]. We observed that when surface iodinated II-23.D7 cells were treated with a cross-linking agent, there was no association of either the
15 25-26 kD LT- α or the 33 kD LT- β related form with an additional membrane protein. This assay demonstrated that receptor binding is not the mechanism by which LT- α and LT- β remain associated with the cell membrane. Sequence analysis of LT- β showed no relationship to
20 either of the two TNF-receptor forms [C. Smith et al., Science, 248, 1019 (1990); T. J. Schall et al., Cell, 61, 361 (1990)].

Affinity Chromatography

Further characterization of the LT- β and LT- α
25 proteins on the surface of II-23.D7 cells was obtained through affinity chromatography. We observed that LT- α /LT- β complex on the surface of PMA-treated II-23.D7 cells bound to lentil lectin, indicating a glycoprotein structure for each form. Hence a lentil
30 lectin chromatography step was used as a purification step prior to antisera affinity chromatography. We bound detergent-solubilized PMA-treated II-23.D7 proteins to lentil lectin sepharose and eluted with α -methyl mannoside. We prepared both control IgG and

SUBSTITUTE SHEET (RULE 26)

- 21 -

anti-LT- α -IgG affinity columns to accurately assess those proteins specifically recognized by the anti-LT- α antiserum. We then applied the proteins that bound to lentil lectin to these columns. We observed
5 that low pH elution of the columns led to the release of the LT- β and LT- α proteins from the anti-LT- α affinity column. SDS-PAGE analysis of the eluate closely resembled the SDS-PAGE analysis of immunoprecipitated proteins from surface iodinated PMA-
10 treated II-23.D7 cells. This comparison demonstrated that similar proteins were purified by the two methods.

We observed that during affinity purification, the ~25 kD LT- α form appeared to be cleaved to a 19-20 kD form, or a "des 20" form. The
15 original isolation of natural LT- α from the RPMI 1788 tumor cell line [B. Aggarwal et al., "Primary Structure of Human Lymphotoxin Derived From 1788 Lymphoblastoid Cell Line," J. Biol. Chem., 260, pp. 2334-2344 (1985)] also yielded an N-terminally cleaved 20 kD LT- α form.
20 One of the methionines is lost in this "des-20" natural LT- α form, producing a different CNBr cleavage pattern from the intact molecule. One-dimensional CNBr digests of the affinity-purified LT- α protein demonstrated a cleavage pattern that is consistent with the truncated
25 natural LT- α form, and we concluded that the affinity-purified "des-20" LT- α form probably results from a similar cleavage as observed with the natural LT- α "des-20" form.

We further observed that LT- β generates a
30 doublet upon partial CNBr cleavage. The cleavage pattern generated by the LT- β protein demonstrated that methionine residue(s) were present, and at least one methionine was within 5-20 residues from either the C-or N-terminus (methionine residues within 1-5
35 residues f either end would not be detected when

SUBSTITUTE SHEET (RULE 26)

- 22 -

cleaved using this mapping technique). This pattern suggested that LT- β does not contain the entire known LT- α sequence.

We observed that the LT- β protein is also
5 expressed by antigen-activated primary cytotoxic T lymphocyte clones. Metabolic labelling of these cells followed by immunoprecipitation with anti-rLT- α revealed LT- β along with small amounts of LT- α . These results demonstrated that LT- β is made by primary T
10 cells as well as by the II-23.D7 hybridoma.

Initial purification of the LT- β and LT- α proteins

We purified these LT- α and LT- β proteins using the following general steps. We first added phorbol myristic acetate (PMA) to II-23.D7 cells.
15 After 24 hours we harvested the cells and washed them with cold serum-free RPMI medium. To the chilled cell pellet we added ice-cold lysis buffer (HEPES, NP-40, EDTA, NaCl, and sodium azide) to which benzamidine, phenyl methyl sulfonyl chloride (PMSF), and N-ethyl
20 maleimide (NEM), soybean trypsin inhibitor, pepstatin and aprotinin had been freshly added. We homogenized the cells gently in a Dounce homogenizer and centrifuged the lysate. We centrifuged and collected the supernatant. We loaded the supernatant onto a
25 lentil-lectin sepharose column equilibrated in lysis buffer to which we had added CaCl_2 and MnCl_2 . We washed the column with lysis buffer with CaCl_2 and MnCl_2 and then eluted with lysis buffer containing α -methyl mannoside. We pooled the eluate fractions and
30 loaded directly onto a rabbit nonspecific IgG sepharose affinity column which was directly connected to a rabbit anti-rLT- α sepharose affinity column. We washed both columns with the same lysis buffer with EDTA followed by lysis buffer wherein the NP-40 had been

SUBSTITUTE SHEET (RULE 26)

- 23 -

replaced with MEGA-8 (Octanoyl-N-methyl glucamide, Boehringer-Mannheim). We eluted the washed columns individually with a solution of 5mM MEGA-8, 50mM glycine, NaCl, benzamidine, and EDTA as described in

5 Browning et al., "Lymphotoxin And An Associated 33-kDa Glycoprotein are Expressed On The Surface Of An Activated Human T-Cell Hybridoma," J. Immunol. 147, pp. 1230-1237 (1991). The first fractions following the pH shift were pooled, lyophilized and resuspended

10 in water with SDS, and dialyzed against a solution of HEPES and SDS. We dried the dialyzed fractions on a speed-vac and resuspended in water. We mixed aliquots with Laemmli loading buffer and electrophoresed on SDS-PAGE. We visualized proteins by silver staining.

15 We observed that LT- α epitope(s) are present on the surface of the II-23.D7 T cell hybridoma only following cell activation such as occurs with PMA treatment. In contrast, when present on primary T-cells, PMA treatment leads to loss of the surface

20 antigen. Additionally, we found that rabbit polyclonal antisera to either recombinant LT- α (produced in CHO cells) or natural LT- α (e.g., Genzyme, Boston, Mass.) recognized the LT- α epitope(s).

We have also observed that our antisera

25 recognizing the LT- α /LT- β complex blocks the MLR, whereas a particular monoclonal antibody recognizing a soluble LT- α does not [M. Shalaby et al., J. Immunol., 141, 499 (1988)]. The LT- α /LT- β complex of the invention, therefore, may be a mediator in T cell

30 activation.

The presence of LT- β with LT- α in immunoprecipitates from cell lysates suggested that either LT- β is antigenically related to LT- α or that LT- β is bound to LT- α or both. To address this issue

35 25 kD and 33 kD bands from ³⁵S-methionine labeled cells

SUBSTITUTE SHEET (RULE 26)

- 24 -

were immunoprecipitated with rabbit polyclonal anti-rLT- α serum, eluted from excised gel slices and subjected to reimmunoprecipitation with either anti-rLT- α polyclonal serum or anti-rLT- α mAb. LT- α , but
5 not LT- β , could be immunoprecipitated with either of the anti-rLT- α antibodies, suggesting that LT- β is not antigenically related to LT- α . These observations indicated that LT- β is physically associated with LT- α .

To further investigate the hypothesis that
10 surface LT- α and LT- β form a complex, we performed isoelectric focusing (IEF) experiments under both denaturing and native conditions, the rationale being that if LT- α and LT- β are physically associated, then they should focus as a complex under native conditions
15 but as separate entities under denaturing conditions. The individual isoelectric points (pI's) for LT- α and LT- β were determined by two-dimensional gel analysis (denaturing conditions) (Fig. 9A). LT- α possesses five charged isomers ranging in pI from 6.5 to 7.3, whereas
20 LT- β possesses four charged isomers ranging in pI from 5.5 to 6.0. When focusing was performed under native conditions, however, LT- α and LT- β focused together as a broad band ranging in pI from 6.3 to 7.2 (Fig. 10A, lanes 6-8). Therefore, the migration of LT- β was
25 significantly retarded under native conditions.

Further purification and identification of LT- α and LT- β

We later purified these LT- α and LT- β proteins using the following general steps. We grew
30 II-23.D7 cells in RPMI medium with fetal bovine serum and we harvested the cells from 50 l of RPMI and resuspended them in medium and we added phorbol myristoyl acetate (PMA). After activation for 6 hours, we harvested the cells by centrifugation and washed
35 them with Dulbecco's phosphate buffered saline. We

SUBSTITUTE SHEET (RULE 26)

- 25 -

suspended the final cell pellet in cold lysis buffer and passed the pellet once through a nitrogen cavitator. We centrifuged the lysed cells and discarded the supernatant. We extracted the pellet
5 overnight in lysis buffer with detergent and then centrifuged it again.

We added the supernatant containing the detergent solubilized membranes to affinity resin composed of monoclonal anti-LT- α coupled to Affi-gel
10 (10) and rocked the suspension overnight. We collected the resin into a small column and washed it with HEPES with nonidet P40, and then with the same buffer with 1% w/v MEGA-8, we eluted the bound proteins with MEGA-8 in glycine buffer and neutralized the fractions
15 immediately with Tris base. We determined the presence of LT- β and LT- α in the fractions by SDS-PAGE analysis and silver staining. We pooled fractions containing these proteins and added SDS, and we dialyzed the pool against 0.1x laemmli sample buffer (using multiple
20 changes to remove the MEGA-8 detergent). We lyophilized the dialyzed solution to dryness and resuspended it in 1/10th the original volume of water.

We ran the sample on an SDS-PAGE gel, blotted onto a ProBlot membrane and stained with Coomassie blue
25 dye. We excised the LT- β and LT- α bands and loaded them into a protein sequencer. We obtained the N-terminal sequence by Edman degradation. We found the sequence of the membrane associated LT- α band to exactly match that described for secreted LT- α , i.e.,
30 Leu Pro Gly Val Gly Leu Thr Pro Ser (amino acid No. 1 to 9.) [P. Gray et al., Nature, 312, pp. 121-124 (1984)]. The Edman degradation analysis revealed that the N-terminal portion of the associated LT- β protein included two possible amino acid sequences: Gly Leu
35 Glu Gly Arg Gly Gln Arg Leu Gln or Gly Leu Glu Gly Arg

SUBSTITUTE SHEET (RULE 26)

- 26 -

Leu Gln Arg Leu Gln. Subsequent DNA analysis using more accurate cDNA techniques confirmed that the correct sequence was Gly Leu Glu Gly Arg Gly Gly Arg Leu Gln.

5 In each case where a surface LT- α form was detected, we were also able to detect LT- β (i.e., in PMA-activated II-23.D7, activated CTL clones, and Hut-78 cells constitutively expressing a surface LT- α form). Because LT- α is secreted from transfected CHO
10 cells in the absence of a surface LT- α form, and because the presence of LT- β is associated with surface-bound LT- α , we concluded that LT- β complexes with LT- α to target it to the cell surface. Biochemically, LT- β and LT- α co-migrate on a non-
15 denaturing isoelectric focusing gel, but when the complex is dissociated with urea, the two proteins run separately. [See Figs. 9A, 10A.] These observations have led us to conclude that LT- α and LT- β exist as a complex on the cell surface.

20 Identifying DNA Sequences That Code For
Lymphotoxin- β And Soluble Lymphotoxin- β Peptides

Lymphotoxin- β was purified by immunoaffinity-chromatography as described above. Direct N-terminal sequencing and in situ trypsin digestion followed by
25 reverse phase HPLC resolution of the digested peptides was performed. [See, Abersold et al., "Internal Amino Acid Sequence Analysis Of Proteins Separated By One Or Two Dimensional Gel Electrophoresis After In Situ Protease Digestion On Nitrocellulose," PNAS, 84, 6970-
30 6974 (1987)]. The resulting N-terminal and internal tryptic fragment peptides were then sequenced using conventional methods. The sequencing of the N-terminus and internal peptides, designated as T105, T87/88, T110 and T67 are shown in Fig. 13.

SUBSTITUTE SHEET (RULE 26)

- 27 -

Two antisense 17-mer oligonucleotide probes GTYTCNGGCTCYTCYTC [SEQ ID NO:9] and GTYTCNGGTTCTCYTC [SEQ ID NO:10], designated 1368 and 1369, respectively, were synthesized to match a portion the sequence of peptide T-87/T-88 and radiolabelled with ³²P. Northern analysis showed that the probe designated 1368 hybridized strongly to a 0.9-1.1 kb mRNA band that was strongly induced in II-23.D7 cells that had been pretreated with phorbol ester as previously described.

10 A cDNA library in the vector pCDM8 was constructed from poly A+ mRNA isolated from II-23.D7 cells induced with PMA for 6 hours. The library was screened with the labelled oligomer designated 1368 and positive clones were isolated following washing with 15 3 M tetramethylammonium chloride at 50°C. Several (>16) clones containing 0.8-0.9 kb inserts were subjected to DNA sequence analysis.

Clone pCDM8/LT-β-12 contained the coding sequence of lymphotoxin-β as shown in SEQ ID NO:1. The other clones were identical except for various truncations at the 5' end. The clone 12 cDNA codes for a functional lymphotoxin-β. Using standard primer extension methods, three additional codons encoding the amino acid residues -- MET GLY ALA -- were identified. 25 A termination sequence AATAAA at position 862-867 was found just prior to the 3' poly A tract indicating that the entire 3' end had been identified. The protein coding sequence encodes for 240 amino acids with a calculated unmodified molecular weight of approximately 30 25,390 kDa.

The 5' end of the LT-β DNA sequence identified from the clone 12 cDNA and primer extension, ATGGGGGCACTGGGGCTG [SEQ ID NO:11] reveals three possible start codons (underlined). [See, e.g., 35 M. Kozak, "An Analysis Of Vertebrate mRNA Sequences:

SUBSTITUTE SHEET (RULE 26)

- 28 -

Intimations Of Translational Control," J. Cell. Biol.,
115, 4, 887-903 (1991).] Engineered LT- β polypeptides
and DNA sequences may be derived by cleaving all or
part of this 5' sequence and substituting a single
5 start codon.

This amino acid sequence profile is typical
of a type II membrane protein. Following a short
(maximum 17) amino acid N-terminal "cytoplasmic" domain
there is an extensive stretch of 30 hydrophobic amino
10 acids which presumably acts as a membrane anchoring
domain. No identical sequences were found within the
available databases. There is one cysteine residue in
the extracellular domain and two methionines within the
last C-terminal 17 amino acids. This is consistent
15 with the very limited cyanogen bromide cleavage pattern
exhibited by this protein.

After the sequence of LT- β was determined,
subsequent comparison revealed that LT- β is a type-II
membrane protein with significant homology to TNF, LT- α
20 and the CD40 protein. These polypeptides share four
regions of sequence conservation in the extracellular
domain. See FIG. 14. Such conservation regions are
likely to enable those polypeptides to form complexes
with each other. [See, e.g., M. Eck and S. Sprang,
25 "The Structure Of Tumor Necrosis Factor- α At 2.6Å
Resolution, Implications For Receptor Binding," J.
Biological Chemistry, 264, 29, pp. 17595-17605 (1989);
E. Jones et al., "Structure Of Tumor Necrosis Factors,"
Nature, 338, pp. 225-228 (1989); M. Eck et al., "The
30 Structure of Human Lymphotoxin (Tumor Necrosis
Factor- β) At 1.9-Å Resolution, J. Biological Chemistry,
267, 4 pp. 2119-2112 (1992); J. Tavernier et al.,
"Conserved Residues Of Tumor Necrosis Factor And
Lymphotoxin Constitute the Framework Of The Trimeric

SUBSTITUTE SHEET (RULE 26)

- 29 -

Structure," Fed. Eur. Biochem. Soc. Lett., 257, 2 (1989)].

Expression of Cloned LT- β

- The pCDM8/LT- β clone 12* or a control
- 5 plasmid, Clone 4 (pCDM8 with a non-functional LT- β cDNA insert), were introduced by electroporation into CHO dhfr⁻ cells and CHO cells stably transfected with human LT- α . After three days, cells were removed with Ca/Mg-free Hank's solution with 5 mM EDTA and stained for
- 10 FACS analysis as described above using either 10 μ g/ml control IgG1 or anti-LT α monoclonal antibody (Boehringer-Mannheim) followed by labelling of bound immunoglobulin with either a FITC or phycoerythrin labelled goat anti-mouse preparation.
- 15 In other experiments, COS cells were electroporated with either clone 4 or clone 12 LT- β cDNA in pCDM8 in the presence or absence of an equal amount of human LT- α cDNA also in the pCDM8 vector and stained for FACS analysis after three days as above.
- 20 Only CHO cells expressing LT- α displayed surface lymphotoxin upon transfection with a functional LT- β DNA, i.e., clone 12.

- Clone 12 lacks an initiating ATG codon, but does possess several CTG initiating codons and hence
- 25 this expression experiment shows that one or several of the 5' CTG codons must initiate translation. CTG codons are known to serve as initiating sites for translation in several eukaryotic proteins [M. Kozak, J. Cell. Biol., 115, 4, 887-903 (1991)]. Similar
- 30 results were observed using the dual transfection system in COS cells, such that only COS cells receiving

* E.coli K12 bearing a plasmid pCDM8/LT- β clone 12, designated BN1289 (MC1061/P3/P33-clone-12) was deposited with ATCC on November 13, 1992.

SUBSTITUTE SHEET (RULE 26)

- 30 -

both LT- α and LT- β DNA displayed substantial surface LT- α in a FACS analysis.

Potential uses of LT- β and LT- α and the LT- α /LT- β complex

5 As noted above, there is considerable structural similarity between LT- β , LT- α , TNF and the CD40 ligand. LT- β , LT- α , TNF and the CD40 ligand are type II membrane proteins and share at least four regions of sequence conservation in the extracellular
10 domain.

In light of this structural similarity, it is of interest that LT- α is found on the surface of activated lymphocytes in a form identical to the secreted molecule but complexed with an additional
15 integral membrane protein presumably anchoring LT- α to the surface. We believe that this unique complex, now determined to be LT- α /LT- β , represents a more relevant form of LT- α and imparts specificity relative to TNF.

The existence of a heterometric complex of
20 lymphokines, while unique to the immune system is reminiscent of signalling molecules in other areas, e.g, the PDGF and inhibin/activin heteromeric complexes. Delineation of the LT- α /LT- β complex poses the possibility of immunoregulatory activities unique
25 to the complex which cannot be mimicked by the LT- α homotrimer. The complex may bind to a unique receptor or receptor chain combinations leading to a high affinity interaction and biologically relevant signalling. The hypothesis of an LT- α /LT- β interaction
30 with a unique receptor complex could account for the relatively poor activity of the LT- α homotrimer relative to TNF in many systems, an observation which cannot be explained by studies on the two known TNF recept rs. [T. Schall et al., Cell, 61, 361-370
35 (1990); C. Smith et al., Science, 248, 1019 (1990)].

SUBSTITUTE SHEET (RULE 26)

- 31 -

The tethering of soluble LT- α to the cell surface via complexation with LT- β indicates that cell-cell contact specific signalling through LT- α /LT- β may be an important aspect of immune regulation. Because the TNF and related LT- α forms are secreted we believe that LT- β may also be secreted. This may be verified in studies using anti-LT- β monoclonal antibodies. Such antibodies may also be used to determine whether LT- β homo-oligomers occur naturally.

10 In general, LT- α and TNF exhibit qualitatively the same spectra of activities, and LT- α and TNF are believed to interact with the same set of receptors (designated the 55 and 80 kD TNF receptors). [C. Smith et al., Science, 248, 1019 (1990); T. Schall et al., Cell, 61, 361 (1990)]. Nonetheless, the quantitative patterns of biological potency exhibited by LT- α and TNF are dramatically different, with LT- α often being much less potent than TNF [see, e.g., Browning et al., J. Immunol., 143, 1859 (1989)]. These observations are difficult to reconcile with the existing receptor binding data. It is possible that the LT- α /LT- β complex imparts unique properties on LT- α such that it now interacts with other as yet undefined receptors. In this case, a LT- α /LT- β complex and the other complexes of this invention would have unique biological properties distinguishing them from either LT- α or TNF. The LT- α /LT- β complex may be used to identify and clone such LT- α /LT- β or LT- β specific receptors. Moreover, further use of the complex may reveal novel biological activities.

Also, while a number of T cell and macrophage cell lines are known to be infectable by the HIV virus, in practice only a small number of cell lines have been useful in propagating the virus in tissue culture. For example, the H9 line, a derivative of Hut-78 originally

SUBSTITUTE SHEET (RULE 26)

- 32 -

exploited by Gallo et al. [M. Popovic et al., Science, 224, 497-500 (1984)], and another human lymphocytic line, C8166, have been valuable for HIV propagation [M. Somasundaran and H. Robinson, Science, 242, 1554-1557 (1988)]. It is possible that surface LT- α expression or the capacity for expression of surface LT- α makes a given cell a good target for HIV proliferation.

A role for TNF has been proposed in enhancing HIV proliferation [L. Osborn et al., Proc. Natl. Acad. Sci. USA, 86, 2336 (1989); Z. Rosenberg and A. Fauci, Immunol. Today, 11, 176 (1990); C. Locardi et al., J. Virology, 64, 5874 (1990); G. Oli et al., Proc. Natl. Acad. Sci. USA, 87, 782 (1990)]. We have found that the II-23.D7 line is infectable with the HIV strain IIIB, but upon PMA treatment the infection by the virus is dramatically increased. The Hut-78 cell line was found to constitutively express a surface LT form, and the C8166 line resembles II-23.D7 in that surface LT appears following PMA treatment. [Ware et al., J. Immunol., article in press (1992)].

Considering these results on the infectability of II-23.D7 by HIV and the relationship between infectable cell lines and surface LT expression, we propose that those lines may be good hosts for HIV infection and replication because the LT- α /LT- β complex and the other polypeptides and complexes of this invention serve a regulatory role. It has been demonstrated that the LT- α gene is induced by expression of the HIV transcriptional activator TAT [K. Sastry et al., J. Biol. Chem., 265, 20091 (1990)] and, moreover, HTLV-1 infection has also been shown to induce LT- α expression [N. Paul, et al., J. Virol., 64, 5412 (1990); E. Tschachler et al., in Human Retrovirology (Raven Press 1990), W. Blattner, eds., p.

SUBSTITUTE SHEET (RULE 26)

- 33 -

105]. Thus, induction of LT- α by HIV infection and consequent LT- α /LT- β complex or other complex expression, or induction of LT- α /LT- β or other complex expression by PMA treatment in cell lines competent to
5 make these proteins, may serve to enhance viral replication. For this reason, antibodies or specific binding proteins (e.g., soluble receptors) to the LT- α /LT- β complex or the other polypeptide complexes of this invention or to soluble forms of those complexes
10 or to LT- β and the other polypeptides of this invention may inhibit HIV proliferation or block HIV-induced T cell death.

Parallels may be drawn between LT- α /LT- β and the CD40 receptor ligand pair where signalling from a T
15 cell surface CD40 ligand provides "help" to the B cell via the CD40 receptor. One could therefore postulate that surface LT- α /LT- β may be a component of T cell regulation of T cells or other cells of hematopoietic lineage such as LAK or NK cells and B-cells. Moreover,
20 this interaction may be dysfunctional in some autoimmune diseases. [See, e.g., R. Watanabe-Fukunaga, Nature, 356, pp. 314-317 (1992).]

Furthermore, a cell surface protein designated as the Fas antigen has been shown to have
25 considerable structural homology with a number of cell-surface receptors including TNF, NGF and the CD40 protein. The Fas antigen has been implicated in mediating apoptosis, a process also referred to as programmed cell death [R. Watanabe-Fukunaga, Nature,
30 356, pp. 314-317 (1992); N. Itoh et al., Cell, 66, pp. 233-243 (1991)]. A strain of mice that demonstrates defects in the Fas antigen develop a systemic lupus erythematosus-like autoimmune disease. This suggests that the structurally similar LT- β r
35 LT- α /LT- β complexes may also play a role in mediating

SUBSTITUTE SHEET (RULE 26)

- 34 -

systemic lupus erythematosus and, therefore, intervention in this pathway may be clinically useful in treating various autoimmune diseases.

- Alternatively, LT- β or an LT- α /LT- β complex may be
- 5 involved in inducing programmed cell death through a cell-cell contact dependent mechanism. The emergence of this family of TNF related ligands to complement the already extensive family of TNF/NGF type receptors suggests the existence of an additional array of
- 10 important regulatory elements within the immune system.

LT- β or a LT- β /LT- α complex may similarly play a role in suppressing the immune system and may be potentially useful in treating allergy and inducing tolerance.

- 15 The location of the TNF/LT locus in the MHC region of the genome has led workers to examine linkage to various autoimmune disease, especially insulin-dependent diabetes melitis. [See, e.g., F. Pociot et al., "A Tumor Necrosis Factor Beta Gene
- 20 Polymorphism In Relation To Monokine Secretion And Insulin-Dependent Diabetes Mellitus," Scand. J. Immunol., 33, 37-49 (1991); K. Badenhop et al., "TNF- α Gene Polymorphisms In type 1 (Insulin-Dependent) Diabetes Mellitus," Diabetologia, 32, 445-448 (1989).]
- 25 Because we found that the LT- β gene is located next to the TNF/LT locus, it is possible that the LT- β gene or its receptor may be involved in this autoimmune condition. Hence, LT- β or its receptor or antibodies to LT- β may comprise a replacement therapy in this form
- 30 of diabetes.

- As discussed above, the LT- β polypeptide, and the polypeptide complexes of this invention, are expected to have a number of potential uses including anti-tumor, T cell activating, or T cell suppressing
- 35 applications, application involving the treatment of

SUBSTITUTE SHEET (RULE 26)

- 35 -

systemic lupus erythematosus, as well as uses in anti-inflammatory compositions and methods. DNA sequences coding for LT- β polypeptides, recombinant DNA molecules including such DNA sequences, and unicellular hosts and
5 animal or human cells in culture transfected with such recombinant DNA molecules may then be employed to produce large amounts of the polypeptides of this invention, substantially free from other human proteins, for use in the compositions and therapies
10 noted above.

Lymphocytes expressing on their surfaces the polypeptide complexes of this invention, and preferably an LT- α /LT- β complex, represent a subset of lymphocytes that may have enhanced abilities to kill tumor cells.
15 As such, this subset would be useful in LAK (lymphokine-activated killer) cell or TIL (Tumor Infiltrating Lymphocyte) cell therapies. [H. Thomas, K. Sikora, "Biological Approaches to Cancer Therapy," Jour. Int. Med. Res., 17, 191 (1989)]. TIL
20 immunotherapy may be improved by gene transfer techniques. Recombinant genes for LT- β and related polypeptides based thereon will be useful therapeutically, for example in TIL therapy, where a LT- β gene, either with or without an LT- α gene, is
25 introduced into T cells isolated from a tumor and introduced to the patient. More preferably, the cells are taken from the patient, transfected with a DNA sequence encoding on expression a polypeptide of this invention, before or after that transfection incubated
30 with a lymphokine, preferably IL-2, and returned to the patient. The transfected T cells (now expressing LT- β and also consequently complexing LT- α) home in on the tumors from which they were removed, where the tumorcidal action of LT- α is delivered directly to the
35 tumors. Likewise, it is contemplated that a LT- β gene

SUBSTITUTE SHEET (RULE 26)

- 36 -

introduced into LAK cells would increase the number of surface complexes on the cells and enhance their activity. Alternatively, introduction of the LT-8 gene into a patient's tumor cells may be useful in creating
5 a tumor vaccine in which the LT-8 modified tumor would trigger an enhanced immune response to the tumor itself. [See, e.g., W.F. Anderson, Science, 256, 808-813 (1992)].

Antibodies or antibody derivatives to the
10 polypeptides and polypeptide complexes of this invention are also useful in conventional immunological methods, e.g., panning or flow cytofluorometric sorting, to enrich for this cell population. [L.J. Wysocki and V.L. Sato, "Panning for Lymphocytes: A
15 method for Cell Selection," PNAS 75, 2844 (1978)].

It is also contemplated that the polypeptides and polypeptide complexes of this invention, or fragments or derivatives thereof, will be useful in cell regulatory or therapeutic applications similar to
20 those in which lymphotoxin- α and tumor necrosis factors are used.

The compositions of this invention will be administered at an effective dose to treat the particular clinical condition addressed. Determination
25 of the particular dose for a given application is well within the skill of the art taking into consideration, for example, the condition and weight of the patient, the extent of desired treatment and the tolerance of the patient for the treatment. Administration of the
30 complexes and polypeptides of this invention, or perhaps peptides derived or synthesized therefrom or using their amino acid sequences, including isolated and purified forms of the polypeptides, or their salts or pharmaceutically acceptable derivatives thereof, may
35 be via any of the conventionally accepted modes of

- 37 -

administration of agents which exhibit anti-tumor, T cell-activating, T cell-suppressing or anti-inflammatory activity.

The compositions used in these therapies may also be in a variety of forms. These include, for example, solid, semi-solid and liquid dosage forms, such as tablets, pills, powders, liquid solutions or suspensions, suppositories, injectable and infusible solutions, and genetic therapy. The preferred form depends on the intended mode of administration and therapeutic application. Modes of administration may include oral, parenteral, subcutaneous, intravenous, intralesional or topical administration. The compositions also will preferably include conventional pharmaceutically acceptable carriers and may include other medicinal agents, carriers, genetic carriers, adjuvants, excipients, etc., e.g., human serum albumin or plasma preparations. Preferably, the compositions are in the form of a unit dose and will usually be administered one or more times a day.

The following are examples which illustrate the LT- β and the LT- α /LT- β complex of this invention and the methods used to characterize them. These examples should not be construed as limiting: the examples are included for purposes of illustration and the present invention is limited only by the claims.

EXAMPLES

We used the following experimental procedures in the examples:

30 Antisera

Recombinant human LT- α (rLT- α) was expressed and secreted by a stably transfected chinese hamster ovary (CHO) cell line into serum-free conditioned

SUBSTITUTE SHEET (RULE 26)

- 38 -

media. We purified the secreted rLT- α from the serum-free conditioned media by a series of Sepharose S, lentil lectin-sepharose and FPLC Mono Q column chromatography steps. The properties of the CHO cell-derived rLT- α preparation have been described. [J. Browning et al., J. Immunol., 143, 1859 (1989)]. We immunized two rabbits (4 and 5) by a lymph node procedure [M. Sigel et al., "Production Of Antibodies By Inoculation Into Lymph Nodes," Met. Enz., 93, 3 (1983)] with 25 μ g of native rLT- α in complete Freund's adjuvant. A third rabbit (6) was immunized via the same route with 25 μ g of denatured rLT- α in complete Freund's adjuvant. We prepared denatured rLT- α by SDS-PAGE followed by electroelution into 0.1% SDS-carbonate buffer.

Using the above methods, three anti-rLT- α antisera were generated, two directed against native rLT- α and a third against SDS-denatured rLT- α . The antisera raised by immunization with native protein (rabbits 4 and 5) could neutralize a 50 unit/ml solution at a dilution of 1:2000-5000. The serum raised against denatured rLT- α (rabbit 6) lacked neutralizing titer, but was weakly reactive with rLT- α on a Western blot. None of the antisera could neutralize r-human TNF nor could they recognize r-human TNF bound to an ELISA plate except for a very weak titer in the antiserum from rabbit 6. Only antiserum from rabbit 6 was capable of recognizing rLT in a western analysis.

We immunized a fourth rabbit with recombinant human TNF. We prepared the polyclonal anti-rTNF rabbit serum via a classical immunization scheme using recombinant human TNF (E.coli derived [D. Weir et al., Handbook Of Experimental Immunology In Four Volumes, Chapter 8 "Immunization Of Experimental Animals"]) in

SUBSTITUTE SHEET (RULE 26)

- 39 -

complete Freund's adjuvant followed by a boost in incomplete Freund's adjuvant. The serum raised against rTNF by immunization had a good neutralizing titer. A neutralizing monoclonal antibody to TNF has been described [Liang et al., "Production And Characterization Of Monoclonal Antibodies Against Recombinant Human Tumor Necrosis Factor/Cachectin," Biochem. Biophys. Res. Comm., 137, 847 (1986)]. Pre-immunization serum was collected from all animals for use as controls.

Cell Growth and T cell Activation

All cells were obtained from the American Type Culture Collection (ATCC), except for the LT- α transfected Chinese hamster ovary (CHO) line that was described previously [Browning, J. Immunol., 143, pp. 1859-1867 (1989)].

Cells were grown in RPMI 1640 supplemented with 1% glutamine, 10 mM HEPES buffer, pH 7.5, penicillin/streptomycin and 10% fetal bovine serum (Hyclone-defined) (designated "complete RPMI"), except for the transfected CHO cells which were grown in Dulbecco's modified Eagle's medium supplemented as above. The human T cell hybridoma, II-23, was a result of a fusion of the human CEM tumor line with activated peripheral T lymphocytes and was further subcloned (II-23.D7) [C. Ware et al., "Human T Cell Hybridomas Producing Cytotoxic Lymphokines: Induction of Lymphotoxin Release And Killer Cell Activity By Anti-CD3 Monoclonal Antibody Or Lectins And Phorbol Ester," Lymph. Res., 5, 313 (1986)]. Human peripheral blood lymphocytes (PBL) were drawn into heparinized glass tubes, isolated by Ficoll-Hypaque centrifugation, washed and resuspended in complete RPMI medium. We treated PBL at 2×10^6 cells/ml with a 1:500 dilution

- 40 -

of OKT3-conditioned medium (~2 ng/ml final) in the presence of 1 µg/ml indomethacin and, in some experiments, with 10 ng/ml rIL-2 (Biogen, Inc., Cambridge, MA)). Human CTL-clones were generated as described [L. Green et al., "Cytotoxic Lymphokines Produced By Cloned Human Cytotoxic T Lymphocytes," J. Immunol., 135, 4034 (1985)] and activated either with irradiated stimulator cells (antigen) or a combination of anti-CD2 monoclonal antibodies (T11₂ + T11₃) provided by E. Reinherz.

Flow Cytometry

We resuspended cells in RPMI 1640 medium with 10% fetal bovine serum (FBS), 0.1% sodium azide and 0.1 mg/ml human IgG at 0°C. Following preincubation with the human IgG, we added additional media containing the desired antisera. Typically the cells were incubated with a final dilution of the anti-rLT-α and anti-rTNF sera of 1:200 for 60-90 min. We washed cells twice with Dulbecco's phosphate buffered saline (PBS) and then incubated them with a 1:500 dilution of fluorescein-labeled goat anti-rabbit IgG (Cappel Durham, N.C.) in the above medium for a minimum of 60 min. Cells were then washed once and either analyzed directly or, in some cases, analyzed following fixation for 10 min. at 0°C with 0.5% paraformaldehyde. We performed two color analyses as above, except that we added phycoerythrin labeled leu-4, leu-2, leu-M3 or leu-16 or leu-19 (Becton-Dickinson, Mountain View, CA) at the second antibody stage. The comparison of surface-bound LT-α with IL-2 receptor levels was done with separate single color analyses with fluorescein-labeled anti-IL-2 receptor (CD25) antibody (Becton-Dickinson, Mountain View, Ca.). Analyses were performed with a FACStar instrument (Becton-Dickinson).

SUBSTITUTE SHEET (RULE 26)

- 41 -

**Adsorption of Neutralizing Anti-rLT- α
Antibodies by Activated II-23.D7 Cells**

We stimulated II-23.D7 and U937 premonocytic cells at 1×10^6 cells/ml for 8 hours with 10 ng/ml of PMA in complete RPMI medium. We washed the cells (1×10^6) three times in medium and aspirated the supernatant to obtain a dry pellet. The cells were then resuspended in 1 ml of medium containing a 1:1000 dilution of anti-rLT- α serum (from rabbit 4) and incubated on ice for 1.5 hours with mixing. Cells were cleared from the antiserum by centrifugation. We mixed the absorbed antiserum (both pre- and post-immune) with an equal volume (50 μ l) of medium containing 15 U/ml of rLT- α and incubated for 20 minutes at room temperature. The mixtures were diluted serially into medium and added to L929 cells (in 0.1 ml) and incubated a further 24 hours. We assessed cell viability by the MTT assay as described [L. Green et al., "Rapid Colorimetric Assay For Cell Viability: Application To The Quantitation of Cytotoxic And Growth Inhibitory Lymphokines" Jour. Immunol. Meth., 70, 257-268 (1984)].

**35 S-Methionine or 35 S-Cysteine
Metabolic Labelling of T cells**

We transferred cells into either cysteine-free or methionine-free RPMI 1640 supplemented with penicillin/streptomycin, glutamine, 10 mM HEPES pH 7.5, 10% v/v dialyzed FBS and 2% v/v conventional RPMI (cold carrier addition). We adjusted the cell concentration to $2-3 \times 10^5$ cells/ml and added 35 S-methionine or 35 S-cysteine to the appropriate medium to a level of 100-200 μ Ci/ml. In the case of freshly activated PBL, the supernatants were gently removed, and the cells were centrifuged, resuspended in labelling medium and added back to the original adherent population. Following a

SUBSTITUTE SHEET (RULE 26)

- 42 -

12-18 hour labelling period, we washed and lysed the cells as described below. With the PBL, cells were removed by pipetting and the adherent population partially removed by treatment with 5 mM EDTA in PBS.

5 Immunoprecipitations

To 0.2-0.5 ml of labeled cell lysate we added 2-4 μ l of rabbit serum. The sample was left for 1-2 hours at 4°C. We then added a 60 μ l aliquot of a 60-75% suspension of washed Protein A sepharose (Pharmacia, Piscataway, N.J.) and rocked the sample for 6-18 hours at 4°C. We washed the Protein A sepharose pellets 3 times with 1% NP-40 in calcium/magnesium free PBS and resuspended them in 50 μ l of Laemmli SDS loading buffer. Typically a single lysate sample was cycled through sequential immunoprecipitations with preimmune anti-rLT- α serum, anti-rTNF antiserum and finally post immune anti-rLT- α antiserum. In one set of experiments, we added 5 mM CaCl_2 and MnCl_2 to the lysate and the lysate was rocked overnight with 75 μ l of 75% suspension of washed lentil lectin-sepharose. The sepharose was washed twice with NP-40/PBS and then eluted with 3 consecutive additions of 75 μ l of 1% NP-40/PBS with 0.25 M α -methyl mannoside. We subjected the pooled washes to the immunoprecipitation protocol.

25 Rabbit anti-rLT Affinity Column

We purified the immunoglobulin fraction from the anti-rLT serum (from rabbit 4) using Protein A sepharose with acid pH elution. The eluted IgG-containing fractions were dialyzed against PBS and concentrated by amicon filtration. The anti-rLT- α -IgG solution (15 ml of 6 mg/ml) was coupled to 8 ml of Affi-g 1 10 resin (Biorad, Richmond, Ca.) as per instructions. We prepared an identical affinity column

- 43 -

using nonspecific rabbit IgG (Cappel, Durham, N.C.). Both columns were washed with PBS, 1 M acetate pH 3.0 with 1% NP-40 and finally with lysis buffer lacking protease inhibitors.

5. Initial Purification of LT- β and LT- α

- We grew II-23.D7 cells (15 l) to a density of 5×10^5 cells/ml and added phorbol myristic acetate (PMA) to give a final concentration of 25 ng/ml. After 24 hours, we harvested the cells and washed them into cold serum-free RPMI medium. To the chilled cell pellet containing 7×10^9 cells we added 100 ml of ice-cold lysis buffer (50 mM HEPES pH 7.5, 1% v/v NP-40, 2 mM EDTA, 0.15 M NaCl and 0.1% sodium azide) to which 5 mM benzamidine, 1 mM phenyl methyl sulfonyl chloride (PMSF) and 0.25 mM N-ethyl maleimide (NEM), 10 μ g/ml soybean trypsin inhibitor, 0.7 μ g/ml pepstatin and 10 μ g/ml aprotinin had been freshly added. We gently homogenized the cells in a dounce homogenizer and centrifuged the lysate at 10,000 x g for 10 minutes. We centrifuged the supernatant at 60,000 x g for 90 minutes and collected the supernatant.

- To the supernatant from the high speed centrifugation we added 5mM CaCl_2 and 5mM MnCl_2 . The supernatant was then loaded onto a 20 ml lentil-lectin sepharose column (Pharmacia, Piscataway, N.J.) equilibrated in lysis buffer plus CaCl_2 and MnCl_2 . We washed the column with lysis buffer (with CaCl_2 and MnCl_2) and then eluted the column with lysis buffer containing 0.25 M α -methyl mannoside.

- We pooled the lentil lectin eluate fractions to give a volume of 50 ml and loaded them directly onto a 2 ml rabbit nonspecific IgG sepharose affinity column. We connected this column directly to a 2 ml rabbit anti-rLT- α sepharose affinity column. We washed

SUBSTITUTE SHEET (RULE 26)

- 44 -

both columns with the same lysis buffer with EDTA, followed by lysis buffer wherein the 1% NP-40 had been replaced with 1% w/v MEGA-8 (Octanoyl-N-methyl glucamide, Boehringer-Mannheim, Indianapolis, IN.).

- 5 We eluted the washed columns individually with 1% MEGA-8, 50 mM glycine pH 2.5, 0.05 M NaCl, 5 mM benzamidine, and 2 mM EDTA. We pooled the first 20 ml following the pH shift, lyophilized the pool and resuspended it in 1 ml of water with 0.05% SDS, and
- 10 dialyzed it against 10 mM HEPES pH 7.5, 0.05% SDS and 0.1% MEGA-8. We dried the dialyzed fractions on a speed-vac and resuspended them in 0.15 ml of water. We mixed aliquots with Laemmli loading buffer and electrophoresed them on SDS-PAGE. The LT- β and LT- α
- 15 proteins were visualized by silver staining.

Iodination of II-23.D7 Cell Surface

- Either control or PMA-induced II-23.D7 cells were washed extensively in calcium/magnesium-free PBS, treated with 1 mM PMSF and 0.25 mM NEM and then washed
- 20 twice. To a 12 x 75 mm glass tube that was coated with 50 μ g of iodogen (Pierce) we added 0.3 ml of cells (1×10^7 total) and 1-2 mCi of 125 sodium iodide. Cells were left with periodic swirling for 25 minutes at room temperature, washed 3 times in PBS with 10% FBS and
- 25 resuspended in lysis buffer as described above. We then removed the nuclei with a 2 minute centrifugation in an Eppendorf centrifuge. We then centrifuged the supernatant an additional 15 minutes. The cleared supernatant was subjected to the immunoprecipitation
- 30 protocol.

SUBSTITUTE SHEET (RULE 26)

- 45 -

1-Dimensional CNBr Peptide Mapping

We electrophoresed samples on a 12% acrylamide SDS-PAGE Laemmli system gel for a short distance and excised the appropriate gel sections. We
5 soaked the gel slices for 1 hour in 1.0 ml of 0.1 N HCl, 0.2% 2-mercaptoethanol with 15 μ l of 700 mg/ml fresh CNBr in 90% formic acid. The slices were then removed and washed for 5 minutes with 0.1 M Tris-Cl pH 8.0, 5 min with 25 mM Tris-Cl pH 8.0 and finally 10 min
10 with 1x Laemmli SDS-PAGE loading buffer. We loaded the slices onto a 15% SDS-PAGE Laemmli gel with a 12% acrylamide stacking gel. We visualized the peptide bands by silver staining or autoradiography of the dried gel.

15 Reimmunoprecipitation

Reimmunoprecipitation of SDS-PAGE-separated antigens was carried out by excising labeled bands from gels, rehydrating them for 10 minutes in TBS, 0.2% SDS, and then dicing the gel slices into small pieces. The
20 proteins were eluted by incubation in 1 ml TBS, 0.2% SDS for 8 hours at room temperature, with rotation. After elution the gel pieces were removed by centrifugation, and NP-40 was added to the supernatant to a final concentration of 2%. The eluted proteins
25 were then immunoprecipitated as above, and reanalyzed by SDS-PAGE.

Isoelectric Focusing (IEF)

Two-dimensional IEF was performed essentially as described by P. H. O'Farrell [J. Biol. Chem., 250,
30 4007-4021 (1975)]. 125 I-labeled antigens were immunoprecipitated from II-23.D7 cell extracts, and the immunoprecipitated proteins were eluted by heating at 100°C for 5 minutes in 100 μ l O'Farrell sample buffer

- 46 -

containing 9.5 M urea. The eluted proteins were then focused (first dimension) on 14 cm x 3 mm tube gels possessing a 2% final concentration of ampholines (range pH 3-10, Sigma) at room temperature for 16 hours at a constant voltage (400 V). The second dimension was 12% SDS-PAGE.

For IEF under native (non-denaturing) conditions, ^{125}I -labeled cell extracts were focused directly on tube gels identical to those above except for the presence of urea. The labeled extract (200 μl volume) was centrifuged at 100,000 x g (30 psi, airfuge) for 10 minutes prior to loading onto the tube gel. The focusing was performed at 4°C under the same conditions as described above. The tube gel was then removed and sliced into 1 cm sections, and the proteins were eluted by incubating each slice in 1 ml TBS, 2% NP-40, 2 mM PMSF for 8 hours at room temperature, with rotation. The supernatants containing the eluted proteins were then immunoprecipitated and analyzed by SDS-PAGE. The pH gradients for both the denatured and native tube gels were determined by measuring the pH's of individual slices from gels run in parallel.

T Cell Proliferation Assays

We isolated and resuspended PBL in complete RPMI as described above except for the substitution of fetal bovine serum with 10% human autologous serum, 1 $\mu\text{g/ml}$ indomethacin and 50 U/ml polymyxin B. In the MLR experiments, autologous serum was the responder's serum. We irradiated stimulator cells from a different donor with 3000 rads. We preheated rabbit sera for 1 hour at 56°C, and diluted and sterile filtered the sera prior to use in proliferation assays. Cells (1×10^5 total) in 0.2 ml in a round bottom 96-well plate were treated with either 5 $\mu\text{g/ml}$ phytohemagglutinin, 1-2

SUBSTITUTE SHEET (RULE 26)

- 47 -

ng/ml OKT3 or $1.5-2 \times 10^5$ irradiated stimulator cells in the presence or absence of various antisera or cytokines. After 3 days (PHA or OKT3 activation) or 5 days (MLR), cells were pulsed with ^3H -thymidine, harvested and counted.

Further Purification of LT- α and LT- β

We grew II-23.D7 cells in RPMI medium with 10% fetal bovine serum and we harvested the cells from 50 ml RPMI medium and resuspended them in medium at a concentration of 4×10^6 cells/ml and we added 50 ng/ml phorbol myristoyl acetate (PMA). After activation for 6 hours we harvested the cells by centrifugation and washed them with Dulbecco's phosphate buffered saline. We suspended the final cell pellet of 4×10^{10} cells in 200 ml of cold lysis buffer (50 mM HEPES buffer, pH 7.0; 0.1 M NaCl, 10 mM EDTA, 5 mM benzamidine, 10 $\mu\text{g/ml}$ each of soybean trypsin inhibitor, aprotinin, chymostatin, leupeptin, antipain, 1 $\mu\text{g/ml}$ pepstatin and 1 mM phenylmethyl sulfonyl fluoride) and passed the pellet once through a nitrogen cavitator. We centrifuged the lysed cells at 40,000 rpm for 60 minutes in a 50.2 Ti rotor and discarded the supernatant. We extracted the pellet overnight in 120 ml of lysis buffer with 1% w/v Nonidet P40 detergent and then centrifuged it as above.

We added the supernatant containing the detergent solubilized membrane proteins to 2 ml of affinity resin composed of monoclonal anti-lymphotoxin (anti-tumor necrosis factor- β from Boehringer Mannheim) coupled to Affi-gel 10 (BioRad) and rocked the suspension overnight. We collected the resin into a small column and washed it with 50 mM HEPES, pH 7.0 with 1% Nonidet P40, and then with the same buffer with 1% w/v MEGA-8 (Boehringer Mannheim). We eluted the

SUBSTITUTE SHEET (RULE 26)

- 48 -

bound proteins with 1% MEGA-8 in 50 mM glycine buffer pH 2.5 and the fractions immediately neutralized with Tris base. We determined the presence of p33 and LT in the fractions by SDS-PAGE analysis and silver staining.

5 We pooled fractions containing these proteins and added SDS to a final concentration of 0.1% w/v and we dialyzed the pool against 0.1x Laemmli sample buffer (multiple changes to remove the MEGA-8 detergent). We lyophilized the dialyzed solution to dryness and

10 resuspended it in 1/10th the original volume of water. We ran the sample on an SDS-PAGE gel, blotted onto a ProBlot membrane (Applied Biosystems) and stained with coomassie blue dye.

This scheme allows one to purify LT- β to a

15 band on a blot. It should be possible for anyone skilled in the art to separate the proteins eluted from the affinity resin by ion exchange chromatography. For example, the complex can be dissociated with urea and the LT- α and LT- β proteins can be separated by, e.g.,

20 MONO Q FPLC (Pharmacia) anion exchange chromatography in Tris-Cl buffer pH 8.0 with 1% nonionic detergent (e.g., MEGA-8, Boehringer-Mannheim) and urea, using a salt gradient elution. This chromatographic technique separates on the basis of differing charges on the

25 proteins. The two proteins are separable in an isoelectric focusing experiment (see, supra) on the basis of charge differences, wherein urea is used to dissociate the LT- α /LT- β complex. Such a combination of affinity chromatography, dissociation in

30 urea/nonionic detergent and ion exchange chromatography allows purification of soluble LT- β or the LT- α /LT- β complex.

SUBSTITUTE SHEET (RULE 26)

- 49 -

Peptide Sequencing Assays

We excised the LT- β and LT- α bands from the Problot and loaded them into a protein sequencer. We obtained N-terminal sequence information by Edman degradation with a model 470A Applied Biosystems sequencer coupled to a 120A PTH amino acid analyzer. LT- β was purified by immunoaffinity-chromatography as described above and tryptic fragment sequence was obtained. [See Abersold et al., "Internal Amino Acid Sequence Analysis Of Proteins Separated By One Or Two-Dimensional Gel Electrophoresis After In Situ, Protease Digestion On Nitrocellulose," PNAS, 84, 6970-6974 (1987)]. That is, protein on the blot was digested with trypsin in situ followed by reverse phase HPLC resolution of the digested peptides. The resulting N-terminal and internal tryptic fragment peptides were then sequenced by Edman degradation. The sequencing of the N-terminal and internal peptides designated as T105, T87/88, T100 and T67 are shown in Fig. 13.

20 Construction of Oligonucleotide Probes

From the sequence of T87/88 the following antisense probes were designed:

1368 GTYTCNGGCTCYTCYTC [SEQ ID NO:9]

1369 GTYTCNGGTTCYTCYTC [SEQ ID NO:10]

25 and synthesized by standard methods. [See, e.g., J. Sambrook et al., Molecular Cloning, A Laboratory Manual, 2ed. (1989)].

Preparation Of An Induced II-23 cDNA Library

We prepared a cDNA sublibrary as follows:

30 We stimulated II-23.D7 cells for six hours with 50 ng/ml PMA to ensure the presence of LT- β mRNA. We isolated the mRNA from these cells and reverse transcribed it into cDNA using techniques well known to

SUBSTITUTE SHEET (RULE 26)

- 50 -

the art. [B. Seed and A. Aruffo, "Molecular Cloning Of The CD2 Antigen, The T-Cell Erythrocyte Receptor, By A Rapid Immunoselection Procedure," PNAS, 84, 3365-3369 (1987)]. Using standard procedures, we ligated double stranded cDNA to a NotI-BstXI linker/adaptor having the following sequence:

5' GCG GCC GCT TTA GAG CAC A 3' [SEQ ID NO:12]
3' CGC CGG CGA AAT CTC 5'

- 10 We then size-selected the cDNA on a 4.2 ml 5-20% potassium acetate gradient, 2 mM EDTA, 1 µg/ml ethidium bromide, in a Beckman® SW60 Rotor for 3 hours at 50,000 rpm at 22°C according to standard methods. We pooled the cDNA fragments of greater than 500 base
- 15 pairs. Then we prepared the vector, pCDM8 (a gift from Brian Seed (Massachusetts General Hospital)). We digested this plasmid with BstXI. To remove the 400 base pair stuffer fragment we centrifuged the mixture on a potassium acetate gradient, as above, and isolated
- 20 the large fragment. We further purified this fragment by agarose gel electrophoresis, and then ligated the cDNA to the vector. In this way, we created recombinant DNA molecules containing DNA sequences for mRNA expressed in induced II-23.D7 cells. We used
- 25 these plasmids to transform E.coli MC1061 P3. The result was a collection of over 1×10^6 recombinant clones comprising a cDNA library for PMA induced II-23.D7 mRNA.

Screening and DNA Sequencing of Clones

- 30 The pCDM8 II-23.D7 library was screened with ^{32}P labelled oligomer 1368 and positive clones were isolated following washing with 3M tetramethylammonium chloride at 50°C. [J. Sambrook et al., Molecular Cloning, A Laboratory Manual, (1989); Jacobs et al.,
- 35 "The Thermal Stability Of Oligonucleotide Duplexes In

SUBSTITUTE SHEET (RULE 26)

- 51 -

Sequence Independent In Tetraalkylammonium Salt Solutions: Application To Identifying Recombinant DNA Clones", Nucleic Acids Research, 16, 10, 4637-4649 (1988)]. Several clones containing 0.9 kb inserts were
5 subject to dideoxynucleotide DNA sequence analysis [Id.].

Expression of LT- β cDNA

The pCDM8/LT- β clone 12 or a control plasmid, clone 4 (pCDM8 with an irrelevant cDNA insert), was
10 introduced by electroporation into CHO dhfr⁻ and a CHO cell stably transfected with human LT- α . After three days, cells were removed with Ca/Mg-free Hank's solution with 5 mM EDTA and stained for FACS analysis as described above using either 10 μ g/ml control IgG,
15 or anti-LT monoclonal antibody (Boehringer-Mannheim) followed by labelling of bound immunoglobulin with either a FITC or phycoerythrin labelled goat anti-mouse preparation. In other experiments, COS cells were electroporated with either clone 4 or clone 12 LT- β
20 cDNA in pCDM8 in the presence or absence of an equal amount of human LT- α cDNA also in the pCDM8 vector and stained for FACS analysis after three days as above.

Northern Analysis of LT- β Expression

Poly A⁺ RNA was isolated from either II-23.D7
25 cells or peripheral blood mononuclear cells (PMBC) using the Fast-Track[™] system provided by Invitrogen. Northern blots were prepared using 2 μ g/lane of RNA and electrophoresis on a formamide gel essentially as described in J. Sambrook et al., Molecular Cloning, A
30 Laboratory Manual, (1989), followed by transfer onto Gene Screen nylon membrane and UV crosslinking. Blots were probed with random primed BstEII/Xmn-I fragment of the LT- β cDNA which had been gel purified, or a

SUBSTITUTE SHEET (RULE 26)

- 52 -

fragment of human LT- α or actin. II-23.D7 cells were induced with 50 ng/ml PMA for varying times and both LT- α and LT- β expression was found to be induced. PBMC were either cultured in RPMI medium alone or in the presence of 1000 units/ml of IL-2 or with OKT3 to activate the T-cells.

Determination of the 5' End of LT- β

The 5' mRNA sequence was determined by primer extension analysis. [B. Wallner et al., Nature 320, 77-81 (1986).] Primer extension using an oligonucleotide primer (probe 360-121 5'GACAGTGATAGGCACCGCCAGCAACAA -3') [SEQ ID NO:13] yielded a roughly 128-130 bp product that upon sequencing using Maxam and Gilbert methodology [A. Maxam and W. Gilbert, "Sequencing End-Labeled DNA With Base-Specific Chemical Cleavages," Methods In Enzymology, 65, 499 (1988)] showed the transcriptional start site to be 7-9 bp upstream of the methionine ATG. The expression exhibited by clone 12 in transient experiments indicates that one or both of the Leu-4 or Leu-6 start site is functional. To verify the 5' mRNA sequence, a cosmid clone, 031A [Spies et al., Science 243, 214 (1989)], was digested with several restriction enzymes, electrophoresed, blotted and probed with a BST E2/Xmn-1 fragment of the LT- β cDNA. The cosmid contained the LT- β gene within a 6 kb EcoR1 fragment which was subcloned into a pUC derivative called pNN 109 which contained a kanamycin resistance gene. Dideoxynucleic acid sequencing gave the entire genomic sequence.

SUBSTITUTE SHEET (RULE 26)

- 53 -

Example 1T cells Express LT-related
Epitopes on Their Surfaces

Under the conditions described above, we
5 activated human peripheral mononuclear cells (PMN) with
OKT3 monoclonal antibody and, after two days in
culture, we analyzed them for expression of LT- α /LT- β
complex related forms using flow cytofluorometric
analysis. In one experiment, the results of which are
10 shown in Figure 1, we cultured fresh PBL for 3 days
with OKT3 and IL-2 and stained them with a 1:200
dilution of antisera to native rLT- α ("LT-4" and "LT-5"
panels on Figure 1, from rabbits 4 and 5 respectively),
denatured rLT- α ("LT-6" panels on Figure 1, from rabbit
15 6) and native rTNF ("TNF" panel on Figure 1, from
rabbit 7). We stained cells with postimmune serum
(solid lines on Figure 1 panels) or with preimmune
serum from each animal (dotted lines on Figure 1
panels). Figure 1 shows that only anti-rLT- α sera from
20 rabbits 4 and 5 recognized epitopes on the activated
peripheral T cells.

In the experiment shown in Figure 2, we
treated II-23.D7 cells with or without 10 ng/ml PMA for
15 hours and stained them as described in the Figure 1
25 experiment, with rabbit 4 anti-rLT- α postimmune serum
(solid line on Figure 2 panels) or with rabbit 4
preimmune serum (dotted line on Figure 2 panels). As
shown in Figure 2, we found that the T cell hybridoma
II-23.D7, which synthesizes LT- α upon phorbol ester
30 (PMA) stimulation, expressed surface LT-related
epitopes upon PMA activation.

To establish that the LT- α -related epitopes
on T cells were related to LT and not to some
c ntaminant in the CHO-cell derived recombinant LT- α
35 pr paration, we treated a 1:1000 dilution of antiserum

SUBSTITUTE SHEET (RULE 26)

- 54 -

from rabbit 4 with PMA-activated, washed II-23.D7 or U937 cells. In the experiment shown in Figure 3, we treated a 1 ml sample of anti-rLT- α sera (1:1000 anti-LT-4) with either no cells (- -), 1×10^8 U937 cells (-O-), 1×10^8 (- -) PMA-activated II-23.D7 cells or 1×10^7 (- -) PMA-activated II-23.D7 cells. We added dilutions of absorbed antisera to a limiting amount of rLT- α in a L929 cytotoxicity assay such that a 1:4000 final dilution was present in the first well. This assay measures the ability of LT- α to kill a mouse fibroblast cell line, L929, within a 24 hour period [L. Green, J.L. Reade, C.F. Ware, "Rapid Colorimetric Assay for Cell Viability: Application to the Quantitation of Cytotoxic and Growth Inhibitory Lymphokines," J. Immunol. Methods, 70, 257 (1984)]. After 24 hours, we assessed cell viability using a MTT readout. Plotted on Figure 3 is optical density (which is proportional to cell viability) vs. the dilution of absorbed antisera. Data represent the average of duplicate wells and duplicates generally were within the range defined by the symbol. As shown in Figure 3, analysis of the neutralizing titer of the absorbed antisera in the standard L929 cytotoxicity assay demonstrated that the activated II-23.D7 cells removed the LT- α neutralizing antibodies, whereas U937 cells were ineffective. These data indicate that the antigenic structures on the membrane surface are actually related to LT- α .

We subjected the hybridoma II-23.D7 to a number of further treatments to examine a number of trivial explanations for the apparent existence of LT- α related epitopes on T cell surfaces. First we ruled out the possibility that LT- α :antibody complexes in the antisera could bind to TNF/LT- α receptors on the hybridoma. Both TNF and LT- α have trimeric structures

SUBSTITUTE SHEET (RULE 26)

- 55 -

which could allow for the presence of antibody binding epitopes within the complex. However, prior saturation of the cellular TNF receptors with soluble TNF or LT- α had no effect on the surface staining. Such saturation
5 should have prevented such an immune complex from binding to such a receptor.

A pH 3 lactic acid treatment, which can release bound TNF from its receptor, had no effect on the signal, suggesting that the LT- α is not receptor
10 bound. However, experiments utilizing ^{125}I -LT- α binding to II-23.D7 cells indicated that receptor bound LT- α was more difficult to remove from its receptor at acidic pH's than TNF.

Mild trypsinization of the cells prior to
15 staining led to a loss of the signal, indicating that the epitope is a protein. To determine whether surface-associated LT- α was phosphatidylinositol linked, the cells were treated with a phosphatidylinositol specific phospholipase C. Under
20 conditions where a PI-linked antigen, LFA-3, could be released [A. Peterson et al., "Monoclonal Antibody And Ligand Binding Sites Of The T Cell Erythrocyte Receptor (CD2)," Nature, 329, 842 (1987)], no effect was observed on the LT- α epitope.

25 We could not stain CHO cells stably transfected with the LT- α gene, either with or without prior PMA activation, indicating that antibodies to CHO derived contaminants in the original rLT- α used to immunize the rabbits were not present in sufficient
30 amounts to contribute to the staining of II-23.D7 cells. Likewise, antibodies generated against any fetal bovine serum proteins contaminating the LT- α preparation would be ineffective in staining T cells since the staining was performed in 10% fetal calf
35 serum.

SUBSTITUTE SHEET (RULE 26)

- 56 -

Pretreatment of the anti-rLT- α serum with rLT- α blocked the staining of LT- α forms on II-23.D7 cells whereas pretreatment with rTNF did not.

Example 2

5 Immunoprecipitation of LT- α -related Proteins on the T cell Hybridoma II-23.D7

We surface iodinated PMA activated II-23.D7 cells and lysed and solubilized the cells in detergent. Immunoprecipitation and SDS-PAGE analysis of the
10 labeled membrane proteins showed that two proteins were recognized by anti-rLT- α antisera. Figure 4A shows the results of SDS-PAGE analysis of the iodinated surface proteins precipitated with either pre-immune (PRE) or post-immune (POST) anti-rLT- α serum (from rabbit 4).

15 As shown in Figure 4A, we observed a 25-26 kD molecular weight form ("LT- α ") that correlated with the expected size of LT- α , and we also saw an additional form of approximately 33 kD ("LT- β or p33"). Neither the preimmune serum from the same rabbit (Figure 4A
20 column PRE) nor anti-rTNF rabbit serum were able to immunoprecipitate any bands from the iodinated, PMA-activated II-23.D7 cells.

1-D partial CNBr peptide mapping of the iodinated bands showed that the 25-26 kD form was
25 cleaved in a pattern identical to that of iodinated rLT- α , thus identifying this band as LT- α . In the experiment shown in Figure 4B, the 25-26 kD and 33 kD bands from panel A were excised, subjected to limited CNBr cleavage and electrophoresed on a SDS-PAGE system.
30 For comparison, cleavages of both rTNF and rLT- α performed in parallel are shown in Figure 4B. The gels were visualized by autoradiography. Lane 1 represents rTNF, lane 2 represents rLT, lane 3 represents LT, and lane 4 represents LT- β . The increased sizes of the

SUBSTITUTE SHEET (RULE 26)

- 57 -

CNBr fragments reflect the increased amount of carbohydrate on natural LT- β . The iodinated 33 kD form was not cleaved by CNBr (lane 4), indicating that it is different from the known LT- α gene product. rTNF was not cleaved with CNBr (lane 1) due to the absence of methionine in this protein.

We undertook metabolic labelling with ^{35}S -methionine and ^{35}S -cysteine coupled with immunoprecipitation to characterize further these LT- α -related surface forms. In the case of the TNF/LT- α pair, the distribution of cysteine and methionine allows one to distinguish both between TNF and LT- α and between forms with and without their signal sequences, as was exploited in studies on the membrane TNF form [M. Kriegler et al., *Cell*, 53, pp. 45-53 (1988)]. In the case of the fully processed cytokines, i.e., secreted forms, TNF contains cysteine and not methionine, while LT- α contains only methionine and not cysteine. LT- α , however, has two cysteine residues in the signal sequence domain and TNF contains a methionine residue in this N-terminal region. Separate cultures of II-23.D7 hybridoma cells were labeled with either ^{35}S -methionine or ^{35}S -cysteine and immunoreactive proteins were precipitated. In an experiment, the results of which are shown in Figure 5, II-23.D7 cells were activated with 10 ng/ml PMA and simultaneously labeled for 8 hours with either ^{35}S -methionine or ^{35}S -cysteine. Both the medium and lysed cells were subjected to consecutive immunoprecipitations with preimmune (rabbit 4) (P), anti-rTNF (T) and anti-rLT- α (rabbit 4) (L) sera in that order. Figure 5 shows an SDS-PAGE autoradiographic analysis of the immunoprecipitates from either supernatants containing secreted proteins or the washed cells. "s-TNF" marks the ^{35}S -methionine labeled

SUBSTITUTE SHEET (RULE 26)

- 58 -

anti-rTNF immunoprecipitated band from the cells that was putatively assigned as the unprocessed 26 kD form of TNF. "Met" and "Cys" refer to the ³⁵S-labeled amino acid employed. Those lanes containing ³⁵S-cysteine
5 were exposed for longer periods of time than the lanes containing ³⁵S-methionine. In the supernatants from these cells (lanes labeled "secreted"), a 25 kD form of LT ("LT-α") was released following PMA treatments by those cells that were labeled with ³⁵S-methionine, but
10 not by those labeled with ³⁵S-cysteine. This pattern is expected for fully processed, secreted LT-α. Longer exposures showed trace amounts of TNF in the supernatant, and the incorporation of label was as expected for fully processed, secreted TNF. We
15 observed the expression of predominantly LT-α with low levels of TNF also at the mRNA level (Shamansky and Ware, unpublished observation). Analysis of the washed cells (lanes labeled "cellular") showed that both the 25-26 kD LT-α, along with the 33 kD LT-β, were present.
20 The relative amounts of the 25-26 kD and 33 kD forms paralleled those observed using surface iodination. The 25-26 kD surface LT-α form lacked cysteine, indicating processing of the leader sequence. The 33 kD form incorporated both ³⁵S-methionine and
25 ³⁵S-cysteine. Longer exposures (not shown) of the film shown in Figure 5 revealed the presence of an anti-TNF immunoprecipitated band from the cells at about 26-27 kD. The band showed incorporation of both labeled cysteine and labeled methionine. The labelling was
30 stronger with cysteine. Since the cys:met ratio is 4:1 in the 26 kD-TNF form, this labelling pattern confirms the identity of this band.

The presence of LT-β with LT-α in immunoprecipitates from cell lysates suggested that
35 either LT-β is antigenically related to LT-α or that

SUBSTITUTE SHEET (RULE 26)

- 59 -

LT- β is bound to LT- α or both. To address this issue 25 kD and 33 kD bands from ^{35}S -methionine labeled cells were immunoprecipitated with rabbit polyclonal anti-rLT- α serum, eluted from excised gel slices and
5 subjected to reimmunoprecipitation with either anti-rLT- α polyclonal serum or mAb. LT- α , but not LT- β , could be immunoprecipitated with either anti-rLT- α antibodies suggesting that LT- β is not antigenically related to LT- α . These observations indicated that
10 LT- β is physically associated with LT- α . We believe that the 33 kD protein is unrelated antigenically to LT- α and simply co-precipitated with LT- α .

With either surface iodination or metabolic labelling, we were unable to detect either of the known
15 55 or 80 kD TNF/LT- α receptor forms associated with LT- α or TNF. Presumably, this is because the receptors are rapidly lost during activation of T cells.
[C. Ware et al., "Regulation Of The CTL Lytic Pathway By Tumor Necrosis Factor," Cellular Immunity And The
20 Immunotherapy Of Cancer, UCLA Symposia on Molecular and Cell Biology M.T. Lotze and O.J. Finn, Eds. Vol. 135, pp. 121-128 (Wiley-Liss, Inc. New York) 1990].

Example 3

25 Biochemical Characterization of Surface LT-forms on II-23.D7 Cells

We purified the LT-related forms on the surface of PMA-treated II-23.D7 cells using affinity chromatography. Using immunoprecipitation techniques, we had noted that both of LT- β and LT- α bound to lentil
30 lectin sepharose, indicating a glycoprotein structure. We bound detergent solubilized PMA-treated II-23.D7 proteins to lentil lectin sepharose and eluted with α -methyl mannoside prior to affinity purification. We prepared both control IgG and anti-IgG columns to more

SUBSTITUTE SHEET (RULE 26)

- 60 -

accurately assess those proteins specifically recognized by the anti-rLT- α serum. Low pH elution of the columns led to the release of about 100-200 ng of the two LT forms from the anti-rLT- α column.

- 5 Figure 6A reflects SDS PAGE analysis of the proteins eluted from anti-rLT- α affinity column prepared from either pre-immune (PRE) or post-immune (POST) rabbit sera. In Figure 6B, the 33 kD and 20 kD bands from the gel in panel A were excised and
- 10 subjected to limiting CNBr cleavage and electrophoresed on a SDS-PAGE system. For comparison, Figure 6B showed CNBr cleavages of rTNF and rLT- α (CHO-derived) performed in parallel. The gels were visualized by silver staining. SDS-PAGE gels of the eluate resembled
- 15 closely gels of immunoprecipitated, surface iodinated PMA-treated II-23.D7 cells, indicating that similar proteins had been purified.

- During the affinity purification, the 25 kD LT- α form appeared to be cleaved to a 19-20 kD form,
- 20 i.e., it now co-migrated with the intact recombinant CHO cell-derived LT- α . The original isolation of natural LT- α from the RPMI 1788 tumor line also yielded an N-terminally cleaved "des-20" LT- α form. One-dimensional CNBr digests of the affinity purified
- 25 proteins showed the cleaved 20 kD LT- α form to have a CNBr cleavage pattern that presumably reflects the truncated nature of this LT- α form. One of the methionines is lost in the "des-20" LT- α form and hence the cleavage pattern would be different from that of
- 30 the intact LT- α form. The 33 kD protein (LT- β) generated a doublet upon CNBr cleavage, and from this it was estimated that the single methionine must lie within 5-20 residues from either the C- or N-terminus. This cleavage pattern shows that the 33 kD protein is
- 35 significantly different from known LT forms. CNBr

SUBSTITUTE SHEET (RULE 26)

- 61 -

cleavage analysis of the surface iodinated 33 kD protein probably gave a similar result; however, the resolution achievable with iodination was insufficient to visualize the doublet. Staphylococcus V8 digestion of the iodinated rLT- α , rTNF and II-23.D7 LT forms showed the rLT- α and 25-26 kD II-23.D7 LT- α form to be resistant to digestion, confirming the assignment of this protein as LT- α . The 33 kD protein was cleaved into several smaller fragments with the pattern resembling closely that of rTNF.

In Figure 7, immunoprecipitated, surface iodinated proteins were resolved on SDS-PAGE analysis and the surface-associated 25-26 kD protein ("sLT- α ") and the 33 kD protein ("LT- β ") bands were excised. Slices were digested with N-glycanase (N-Gly), with a mixture of neuraminidase and O-glycanase (O-Gly), or with all three enzymes. The digested slices were rerun on SDS-PAGE and an autoradiogram of the dried gel is shown. As shown in Figure 7, immunoprecipitation of iodinated surface LT forms followed by digestion with either or both N- and O-glycanases showed the 25-26 kD LT- α form to contain an N-linked oligosaccharide. The 25-26 kD LT- α form contains only one N-linked site which would correlate well with the size change upon N-glycanase digestion. Likewise, the 33 kD form (LT- β) lost about 3 kD of size upon treatment with N-glycanase, suggesting the presence of one N-linked oligosaccharide. In contrast to the 25-26 kD LT- α form, O-glycanase treatment did not affect the molecular weight of LT- β . The lack of cleavage by a glycanase, however, is not definitive evidence for the lack of a carbohydrate.

SUBSTITUTE SHEET (RULE 26)

- 62 -

Example 4Reimmunoprecipitation of LT- α

The coprecipitation of LT- β with LT- α suggested that these proteins are either antigenically related or physically associated. To address this issue we tested whether or not SDS-PAGE separated LT- α (p25) and LT- β proteins could be immunoprecipitated. LT- α and LT- β labeled with ^{125}I or ^{35}S -Met were first partially purified by immunoprecipitation and separated by SDS-PAGE. The labeled bands were excised, rehydrated in buffer, and the proteins eluted. The eluted proteins were then subjected to a second round of immunoprecipitation using either polyclonal or monoclonal anti-rLT- α antibodies (Fig. 10). Rabbit anti-rLT- α reimmunoprecipitated LT- α ("p25", lane 2) but not LT- β (lane 3). The anti-rLT- α mAb precipitated LT- α (lane 5) and a 21 kD protein ("p21", lane 4), which, as shown below, is a precursor of LT- α ; however, it did not precipitate LT- β (lane 6). The results indicate that after LT- α and LT- β are separated by SDS-PAGE, both polyclonal and monoclonal anti-rLT- α antibodies are capable of reacting with LT- α but not with LT- β . This data provides evidence that LT- β is not antigenically related to LT- α . However, we cannot rule out the possibility that putative LT- β cross-reactive epitopes are lost after denaturation, whereas the LT- α epitopes remain intact.

Figure 8 shows the results of reimmunoprecipitation of ^{125}I -labeled and ^{35}S -Met-labeled p25 and p33 proteins eluted from SDS-PAGE gels. LT- α and LT- β species from ^{125}I -labeled II-23.D7 cells (lanes 2,3) and ^{35}S -labeled cells (lanes 4,5,6) were eluted from gel slices as described above. The elutes were immunoprecipitated with either the anti-rLT- α serum (lanes 2,3) or the anti-rLT- α mAb (lanes 4,5,6), and

SUBSTITUTE SHEET (RULE 26)

- 63 -

the reprecipitated proteins analyzed by SDS-PAGE and autoradiography. Lane 1 is a control lane for the identification of LT- α and LT- β .

Example 5

5 Isoelectric Focusing of LT- α and LT- β

Figures 9 and 10 (each including an autoradiograph (9A, 10A) and a calibration curve graphing migration distance vs. pH (9B, 10B)) depict isoelectric focusing analysis under denaturing (Fig. 9) and native (Fig. 10) conditions. Two-dimensional gel analysis was carried out as described above on ^{125}I -labeled LT- α and LT- β that had been immuno-precipitated from II-23.D7 cell extracts. The 2-D gel analysis was performed under denaturing conditions in the presence of urea (Fig. 9A). In contrast, native IEF was performed in 1% NP-40 without urea. ^{125}I -labeled II-23.D7 cell extract was first focused on a tube gel at 4°. After focusing, the tube gel was cut into 1 cm sections, the focused proteins eluted from those sections, immunoprecipitated, and analyzed by SDS-PAGE (Fig. 10A). Immunoprecipitated material from gel lanes 1-12 correspond to tube gel slices 2-13. pH gradients were generated for both the denatured and native tube gels based on 1 cm gel increments. These are also shown below each autoradiogram as 9B and 10B, respectively. Biochemically, LT- β and LT- α comigrate on a non-denaturing isoelectric focusing gel, but when the complex is dissociated with urea, the two proteins run separately. [See Figs. 9A, 10A] These observations led us to conclude that LT- α and LT- β exist as a complex on the cell surface.

SUBSTITUTE SHEET (RULE 26)

- 64 -

Example 6

Regulation of LT- α Expression

Table I set forth below, summarizes the
results of a survey using flow cytometric
5 analysis of various cell types for the expression of a
surface form of LT- α .

SUBSTITUTE SHEET (RULE 26)

Table I

**Expression of LT- α and TNF Related Epitopes
on the Surfaces of Different Cells**

5	Cell	Treatment	Surface Expression of:	
			LT- α	TNF
10	Peripheral Mononuclear cells	Resting	+	-
		OKT3	++	-
	Leu-4 ⁺ (CD3)	Resting	+	-
		OKT3	++	-
15		PMA	-	nd.
		IL-2	++	nd.
	Leu-2 ⁺ (CD8)	OKT3	++	nd.
	Leu-3 ⁺ (CD4)	OKT3	++	nd.
20	Leu-M3	Resting	-	+
		OKT3/LPS/IFN- γ	-	++
	Leu-19 ⁺ (NK)	IL-2 (LAKs)	++	nd.
	Leu-16 ⁺ (B's)	Resting	+/-	-
25		PWM	+	nd.
	CTL-clones	Control	+/-	-
		PMA	nd.	nd.
		Allogeneic Stim.	++	nd.
30		Anti-T11 2+3	++	-
	T cell Hybridoma (II-23.D7)	Control	-	-
		PMA	++	-
		PMA + A23187	++	-
35	Hut-78	Control	+	-
		PMA	+	-
	C8166	Control	-	nd.
		PMA	+	nd.
35	RPMI-1788	Control	-	-
		PMA	-	-
	rLT-producing CHO cell line	Control	-	-
		PMA	-	-
35	Jurkat		+/-	nd.
	HL-60		-	-
	U937		-	nd.
	Raji		+	nd.
35	K562		+/-	nd.

- 66 -

The most striking observation from these studies was the restriction of surface LT- α expression to T and B cells. Leu-M3, a monocyte marker, and leu-4 (CD3) antibodies were used in two-color flow
5 cytofluorometric analysis to observe each cell population separately. There was an excellent distinction between surface TNF and surface LT- α in this analysis in that monocytes expressed only surface TNF whereas T cells displayed only surface LT- α . This
10 result is shown in Figure 11.

In the experiment depicted in Figure 11, PBL were treated for 8 hours with a mixture of LPS (1 μ g/ml), Interferon- γ (200 U/ml) and OKT3 (1ng/ml) and then stained for LT- α (anti-rLT- α serum from rabbit
15 5) or TNF (anti-rTNF serum from rabbit 7) followed by FITC anti-rabbit labelling. Cells were counterstained with either phycoerythrin-leu-4, a pan T cell marker, or phycoerythrin leu-M3, a monocyte marker. "T cell panels" were gated for leu-4⁺ cells while the
20 "monocyte" panels were gated for leu-M3⁺ cells. Cells were stained with preimmune (dotted lines) or postimmune sera (solid lines). The monocytic tumor lines HL-60 and U937 did not stain for LT- α . By using two color flow cytofluorometric analysis, the T4 and T8
25 subclasses of activated PBL were found to display similar levels of surface-associated LT- α . In general, it appears that primary T-cells capable of expressing LT- α are also capable of displaying surface LT- α form(s).

30 Examination of three different human donors showed that a surface LT- α form was present on freshly isolated, resting peripheral T cells. In the case of PBL, OKT3 activation or simply IL-2 treatment of the cells led to increased expression. By using
35 fluorescence channel numbers, we have attempted to

SUBSTITUTE SHEET (RULE 26)

- 67 -

quantitate both surface LT- α and IL-2 receptor (CD25) expression during OKT3 activation. Maximal surface LT- α induction by OKT3 appeared to precede the peak expression of IL-2 receptor (TAC expression) in the bulk culture, thus the surface-bound LT form (LT- α /LT- β complex) appears to be an early T-cell activation antigen. It was found that both anti-T11 and allogeneic antigen were capable of causing the appearance of LT- α on the surface of cloned cytotoxic T cells. Likewise, PMA stimulation was necessary to induce the appearance of LT- α on the surface of the II-23.D7 hybridoma. It appears that T cell activation increases surface LT- α form(s). Peripheral lymphocytes, in contrast to the II-23.D7 hybridoma, down-regulate surface LT- α form(s) very quickly following PMA treatment. Likewise, in a two-color analysis of OKT3 activated PBL populations, Dr⁺ cells, which should include T cells in advanced stages of activation, lacked surface LT- α form(s).

Activation of fresh PBL with high levels of IL-2 generated lymphokine-activated killer cells (LAK cells). As shown in Figure 12, two color flow cytofluorometric analysis using anti-rLT- α and leu-19, a NK/LAK cell marker, showed LAK cell expression of surface LT- α forms to resemble the T cell hybridoma, II-23.D7. In the experiment depicted in Figure 12, PBL were cultured for 5 days with 20 ng/ml IL-2 and then stained for a two color analysis with phycoerythrin labeled leu-19 and anti-rLT (rabbit 5) as described above. Figure 12 shows surface LT- α levels on leu-19⁺ cells that were stained with preimmune (dotted lines) or postimmune sera (solid lines). Thus, LAK cells appeared to have the highest levels of surface LT- α forms of any primary cell type.

SUBSTITUTE SHEET (RULE 26)

- 68 -

Example 7Functional Relevance of Total TNF or LT- α to
T cell Activation

To examine the functional relevance of TNF
5 and LT- α to the T cell activation process, we included
the rabbit anti-rLT- α and anti-rTNF sera in mixed
lymphocyte response (MLR) and OKT3 activation assays.
MLR is a standard immunological assay which tests the
ability of an individual's T cells to recognize another
10 person's T cells as foreign and respond to their
presence by proliferating. Table II, set forth below,
presents data from MLR experiments using various
responder/stimulator combinations.

- 69 -

TABLE II

Effects of LT and TNF Antibodies
on a 5-Day Mixed Lymphocyte Culture

Cells ^b	Addition	³ H-Thy. (S.D.) % Change ^a cpm x 1000	
Responder A	none	4.7 (0.8)	-
Stimulator B	none	4.8 (0.5)	-
A + B	none	20.3 (3.6)	0%
A + B	r-TNF ^c	28.0 (1.4)	+49%
A + B	r-LT	32.8 (3.2)	+80%
Responder C	none	6.3 (0.8)	-
Stimulator B	none	4.8 (0.7)	-
C + B	none	30.0 (5.4)	0%
C + B	r-TNF	36.9 (6.0)	+28%
C + B	r-LT	36.1 (5.8)	+24%
Responder A	none	5.3 (0.6)	-
Stimulator D	none	1.5 (0.5)	-
A + D	none	22.2 (2.6)	0%
Responder D	none	7.8 (1.0)	-
Stimulator A	none	1.6 (0.3)	-
D + A	none	24.3 (7.0)	0%
		<u>Preimmune</u>	<u>Postimmune</u>
A + B	Anti-LT-4 ^d	26.8 (2.4) +41%	8.6 (0.2) -74%
A + B	Anti-LT-5	27.4 (4.4) +45%	11.0 (1.2) -59%
A + B	Anti-LT-6	23.1 (1.0) +18%	25.7 (5.7) +35%
A + B	Anti-TNF	26.0 (2.2) +36%	12.6 (2.7) -49%
A + B	Anti-TNF mAb	15.1 (2.2) -33%	4.7 (0.7) -99%
C + B	Anti-LT-4 ^d	41.1 (3.5) +44%	20.9 (5.8) -36%
C + B	Anti-LT-5	35.5 (6.9) +22%	17.8 (2.6) -49%
C + B	Anti-LT-6	39.4 (7.9) +38%	39.1 (5.3) +36%
C + B	Anti-TNF	39.8 (4.3) +39%	24.4 (3.2) -22%
C + B	Anti-TNF mAb ^e	37.8 (7.3) +31%	20.6 (1.8) -37%
A + D	Anti-LT-5	28.6 (2.1) +37%	12.5 (2.4) -59%
A + D	Anti-LT-5	32.8 (6.4) +63%	14.5 (3.6) -47%
D + A	Anti-LT-5	28.0 (1.2) +23%	20.8 (1.4) -21%
D + A	Anti-LT-5 ^f	28.1 (2.1) +24%	19.2 (0.7) -31%

See notes below.

^aPercent change refers to the increase or decrease in ³H-thymidine incorporation after correction for a background and f responder cells alone.

SUBSTITUTE SHEET (RULE 26)

- 70 -

^bStimulator cells were irradiated with 3000 rads and are denoted with a "**". Low level proliferation was still evident in stimulator population. The ratio of responder to stimulator was 1/1.5.

5 ^crTNF and rLT were added to a level of 10 ng/ml.

^dAntisera were heat inactivated for 1 hour at 56°C, filtered and used at a final dilution of 1:250.

^eThe monoclonal anti-TNF was a purified mouse IgG_{2a} antibody used at 2 µg/ml and the control in this case
10 was pure mouse UPC 10 (IgG_{2a}).

^fIn these cases, the immunoglobulin fraction was purified from the serum and used at a final concentration of 50 µg/ml.

As shown in Table II, the neutralizing anti-
15 rLT-α sera (rabbits 4 and 5) inhibited the proliferative response as assessed at five days whereas preimmune sera or the non-neutralizing anti-rLT-α (rabbit 6) sera had mild stimulatory effects. As previously reported [M. Shalaby et al., J. Immunol.,
20 141, 499 (1988)], polyclonal and a monoclonal anti-TNF preparations were also inhibitory. These assays were carried out under excess stimulator cell conditions and hence the inhibition may not be optimized. The serum levels employed in Table II are rather high, but in
25 other experiments (data not shown), antibody dilutions up to 1:1000 were still inhibitory. PHA or OKT3 stimulated T cell proliferation was also inhibited to a lesser extent (data not shown). These data indicated that LT or LT-related epitopes on T cell surfaces may
30 be involved in T cell activation.

A previous study using the MLR assay and neutralizing monoclonal antibodies implicated TNF but not LT-α in T cell activation and subsequent proliferation in this system. [M. Shalaby et al., J. Immunol., 141, 499 (1988)]. In that study, monoclonal
35

SUBSTITUTE SHEET (RULE 26)

- 71 -

anti-rLT- α antibodies had no effect on the MLR assay. Our studies indicate that neutralizing polyclonal anti-CHO-cell-derived-rLT- α sera were able to partially inhibit the MLR, suggesting a role for some form of LT

5 in this system. The reasons for this discrepancy are not clear, although there may be some differences in the nature of these antibody preparations. The monoclonal antibodies were generated against

10 glutaraldehyde-cross linked natural LT- α (RPMI 1788 secreted), whereas polyclonal anti-rLT- α sera were prepared using native r-LT- α (recombinant CHO-cell derived) with Freund's adjuvant injected directly into the lymph nodes. The depot effect in the success of the latter system was probably important considering

15 the difficulties reported in immunizing mice [T. Bringman et al., "Monoclonal Antibodies to Human Tumor Necrosis Factors Alpha and Beta: Application For Affinity Purification, Immunoassays, And As Structural Probes," Hybridoma, 6, 489 (1987)]. These data suggest

20 that the blocking effect of our polyclonal anti-rLT- α sera on the MLR is a result of recognition of the surface LT form(s) by the sera rather than recognition of the conventional soluble LT- α form.

Example 8

25 Purification And Initial Sequencing Of LT- α And LT- β

We obtained N-terminal sequence information for LT- α and LT- β by Edman degradation as described above. We found the sequence of the membrane associated LT- α band to be as follows: Leu Pro Gly Val

30 Gly Leu Thr Pro Ser. This sequence matches with the known sequence of secreted LT- α . The Edman degradation analysis revealed that the N-terminal portion of the associated LT- β protein included two possible amino acid sequences: Gly Leu Glu Gly Arg Gly Gln Arg Leu

SUBSTITUTE SHEET (RULE 26)

- 72 -

Gln or Gly Leu Glu Gly Arg Leu Gln Arg Leu Gln.
Subsequent DNA analysis confirmed the former sequence except that glycine was incorrectly identified as glutamine at cycle 7.

5

Example 9Sequencing And Cloning LT-B

- As described above, we obtained sequences for several peptides using the method of Abersold et al. Two antisense 17-mer oligonucleotide probes
- 10 GTYTCNGGCTCYTCYTC [SEQ ID NO:9] and GTYTCNGGTTCTCYTCYTC [SEQ ID NO:10] were synthesized to match a portion of the sequence of one of those peptides, T-87/T-88. Those probes were radiolabelled with ³²P. Northern analysis (as described in J. Sambrook et al., Molecular
- 15 Cloning a Laboratory Manual, 2d ed. (1989)) showed that probe 1368 (GTYTCNGGCTCYTCYTC) [SEQ ID NO:10] hybridized strongly to a 0.9-1.1 kb mRNA band that was strongly induced in II-23.D7 cells which had been pretreated with phorbol ester as previously described. A cDNA
- 20 library in the vector pCDM8 (available from Invitrogen Corporation, San Diego, California) was constructed from poly A+ mRNA isolated from II-23.D7 cells induced with PMA for 6 hours. [See, B. Seed and A. Aruffo, PNAS, 84, 3365-3369 (1987)]. The library was screened
- 25 with labelled oligomer 1368 and positive clones were isolated following washing with 3 M tetramethylammonium chloride at 50°C. [See Jacobs et al., Nucleic Acids Research, 16, 10, 4637-4649 (1988)]. Several clones containing 0.9 kb inserts were subjected to DNA
- 30 sequence analysis. Clone pCDM8/LT-B-12 (clone 12) was found to contain the coding sequence. The other clones were identical except for various 1-30 bp truncations at the 5' end. One potential clone (clone 4) contained a frameshift and was used as a control in transfection

SUBSTITUTE SHEET (RULE 26)

- 73 -

experiments. A termination sequence, AATAAA at position 862 was found just prior to a poly A tract indicating that the entire 3' end had been identified. The identified protein sequence encodes
5 for at least 240 amino acids with a calculated molecular weight of 25,390 and a domain structure typical of a type II membrane protein. The present data suggest that the initiating CTG leads to a processed N-terminus starting with gly 5 (met=1), i.e.,
10 the CTG encoded leucine is either not translated or the leucine-4 residue is further processed yielding the mature N-terminus obtained by amino acid sequencing.

The 33 kDa size of LT- β results from N-linked glycosylation as previously defined and this result is
15 corroborated by the presence of one potential N-linked carbohydrate site in the amino acid sequence at a position identical with a similar site found in the CD40 ligand. A typical N-linked sugar residue can add approximately 3-4 kDa MW, hence, the final molecular
20 weight is close to the observed 30-33 kDa.

No identical sequences were found within the databases. There is one cysteine residue in the extracellular domain and two methionines within the last C-terminal 17 amino acids, in agreement with the
25 very limited cyanogen bromide cleavage pattern exhibited by this protein.

Example 10

Expression Of LT- β

The pCDM8/LT- β clone 12 or a control plasmid,
30 clone 4 (pCDM8 with a non-functional cDNA insert), were introduced by electroporation into CHO dhfr- and a CHO cell stably transfected with human LT- α . After three days, cells were removed with Ca/Mg-free Hank's solution with 5 mM EDTA and stained for FACS analysis

SUBSTITUTE SHEET (RULE 26)

- 74 -

as described above using either 10 µg/ml control IgG1 or anti-LT monoclonal antibody (Boehringer-Mannheim) followed by labelling of bound immunoglobulin with either a FITC or phycoerythrin labelled goat anti-mouse
5 preparation.

In a different experiment, COS cells were electroporated with either clone 4 or clone 12 LT-β cDNA in pCDM8 in the presence or absence of an equal amount of human LT-α cDNA also in the pCDM8 vector and
10 stained for FACS analysis after three days as above. Only COS cells expressing LT-α displayed surface lymphotoxin upon transfection with a functional LT-β DNA, i.e., clone 12.

Clone 12 lacks an initiating ATG codon, but
15 does possess several CTG initiating codons and hence this expression experiment shows that one or several of the 5' CTG codons can initiate translation. CTG codons are known to serve as initiating sites for translation in several eukaryotic proteins. [M. Kozak, J. Cell.
20 Biol., 115, 4 (1991)]. Similar results were observed using the dual transfection system such that only COS cells receiving both LT-α and LT-β DNA displayed substantial surface LT-α in a FACS analysis.

Northern analysis of II-23.D7 cells showed
25 hybridization of the LT-β cDNA to a 0.9-1.0 kb mRNA indicating that the cloned cDNA represents essentially all of the transcribed gene. See Fig. 15. The LT-β gene was expressed at low levels in untreated II-23.D7 hybridoma cells; however, upon cell activation with
30 phorbol ester mRNA levels increased dramatically. See Fig. 16.

SUBSTITUTE SHEET (RULE 26)

- 75 -

Example 11Homology Between LT- β and Other Members of the TNF Family Of Lymphokines

Cloning of the cDNA encoding LT- β revealed
5 that LT- β is a type-II membrane protein with
significant homology to TNF, LT- α and the ligand for
the CD40 receptor. These proteins are known to bind to
members of the TNF/NGF receptor family. LT- β , TNF,
LT- α and the ligand for the CD-40 receptor share four
10 regions of sequence conservation in the extracellular
domain. See Fig. 13. These domains are located on the
face of the TNF and LT- α crystal structures and are
likely to be involved in intersubunit interactions.

Example 1215 Determination of the 5' End of LT- β Reveals Several Possible Start Sites

The 5' mRNA sequence was determined by primer
extension analysis. Primer extension analysis revealed
that the transcriptional start site for the LT- β
20 protein was approximately 7-9 base pairs upstream of
the methionine ATG. Thus, the mRNA possesses at least
3 possible translation start sites, the Met-1, Leu-4
and Leu-6 codons. Transient experiments with clone 12
showed that one or both of the Leu-4 or Leu-6 start
25 sites is functional. The 5' mRNA sequence was verified
by determining the LT- β genomic sequence using a cosmid
clone 031A, described above, and dideoxynucleic acid
sequencing.

SUBSTITUTE SHEET (RULE 26)

- 76 -

SEQUENCE LISTING

(1) GENERAL INFORMATION:

- (i) APPLICANT: BIOGEN, INC.
Reagents of the University of,
California
- (ii) TITLE OF INVENTION: LYMPHOTOKIN-BETA, LYMPHOTOKIN-BETA
COMPLEXES, PHARMACEUTICAL PREPARATIONS AND THERAPEUTIC
USES THEREOF
- (iii) NUMBER OF SEQUENCES: 13
- (iv) CORRESPONDENCE ADDRESS:
 - (A) ADDRESSEE: c/o FISH & NEAVE
 - (B) STREET: 1251 Avenue of the Americas
 - (C) CITY: New York
 - (D) STATE: New York
 - (E) COUNTRY: U.S.A.
 - (F) ZIP: 10020
- (v) COMPUTER READABLE FORM:
 - (A) MEDIUM TYPE: Floppy disk
 - (B) COMPUTER: IBM PC compatible
 - (C) OPERATING SYSTEM: PC-DOS/MS-DOS
 - (D) SOFTWARE: PatentIn Release #1.0, Version #1.25
- (vi) CURRENT APPLICATION DATA:
 - (A) APPLICATION NUMBER:
 - (B) FILING DATE:
 - (C) CLASSIFICATION:
- (vii) PRIOR APPLICATION DATA:
 - (A) APPLICATION NUMBER: US 07/990,304
 - (B) FILING DATE: 04-DEC-1992
- (viii) ATTORNEY/AGENT INFORMATION:
 - (A) NAME: HALEY Jr., James F.
 - (B) REGISTRATION NUMBER: 27,794
 - (C) REFERENCE/DOCKET NUMBER: B129CIP2
- (ix) TELECOMMUNICATION INFORMATION:
 - (A) TELEPHONE: (212) 596-9000
 - (B) TELEFAX: (212) 596-9090
 - (C) TELEX: 14-8367

(2) INFORMATION FOR SEQ ID NO:1:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 726 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: double
 - (D) TOPOLOGY: linear

SUBSTITUTE SHEET (RULE 26)

- 77 -

(ix) FEATURE:

(A) NAME/KEY: CDS

(B) LOCATION: 1..723

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:1:

CIG GGG CIG GAG GGC AGG GGT GGG AGG CTC CAG GGG AGG GGT TCC CTC Leu Gly Leu Glu Gly Arg Gly Gly Arg Leu Gln Gly Arg Gly Ser Leu	48
1 5 10 15	
CIG CTA GCT GIG GCA GGA GGC ACT TCT CIG GIG ACC TIG TIG CIG GCG Leu Leu Ala Val Ala Gly Ala Thr Ser Leu Val Thr Leu Leu Leu Ala	96
20 25 30	
GIG COT ATC ACT GTC CIG GCT GIG CIG GGC TTA GIG CCC CAG GAT CAG Val Pro Ile Thr Val Leu Ala Val Leu Ala Leu Val Pro Gln Asp Gln	144
35 40 45	
GGA GGA CIG GTA ACG GAG ACG GGC GAC CCC GGG GCA CAG GGC CAG CAA Gly Gly Leu Val Thr Glu Thr Ala Asp Pro Gly Ala Gln Ala Gln Gln	192
50 55 60	
GGA CIG GGG TTT CAG AAG CIG CCA GAG GAG GAG CCA GAA ACA GAT CTC Gly Leu Gly Phe Gln Lys Leu Pro Glu Glu Glu Pro Glu Thr Asp Leu	240
65 70 75 80	
AGC CCC GGG CTC CCA GCT GGC CAC CTC ATA GGC GCT CCG CIG AAG GGG Ser Pro Gly Leu Pro Ala Ala His Leu Ile Gly Ala Pro Leu Lys Gly	288
85 90 95	
CAG GGG CTA GGC TGG GAG ACG ACG AAG GAA CAG GGG TTT CIG ACG AGC Gln Gly Leu Gly Trp Glu Thr Thr Lys Glu Gln Ala Phe Leu Thr Ser	336
100 105 110	
GGG ACG CAG TTC TCG GAC GGC GAG GGG CIG GCG CTC CCG CAG GAC GGC Gly Thr Gln Phe Ser Asp Ala Glu Gly Leu Ala Leu Pro Gln Asp Gly	384
115 120 125	
CTC TAT TAC CTC TAC TGT CTC GTC GGC TAC CCG GGC CCG GCG CCC COT Leu Tyr Tyr Leu Tyr Cys Leu Val Gly Tyr Arg Gly Arg Ala Pro Pro	432
130 135 140	
GGC GGC GGG GAC CCC CAG GGC GGC TCG GTC ACG CIG GGC AGC TCT CIG Gly Gly Gly Asp Pro Gln Gly Arg Ser Val Thr Leu Arg Ser Ser Leu	480
145 150 155 160	
TAC CCG GCG GGG GGC GGC TAC GGG CCG GGC ACT CCC GAG CIG CIG CTC Tyr Arg Ala Gly Gly Ala Tyr Gly Pro Gly Thr Pro Glu Leu Leu Leu	528
165 170 175	
GAG GGC GGC GAG ACG GIG ACT CCA GIG CIG GAC CCG GGC AGG AGA CAA Glu Gly Ala Glu Thr Val Thr Pro Val Leu Asp Pro Ala Arg Arg Gln	576
180 185 190	

SUBSTITUTE SHEET (RULE 26)

- 78 -

GGG TAC GGG OCT CTC TGG TAC ACG AGC GTG GGG TTC GGC GGC CTG GIG Gly Tyr Gly Pro Leu Trp Tyr Thr Ser Val Gly Phe Gly Gly Leu Val 195 200 205	624
CAG CTC CCG AGG GGC GAG AGG GIG TAC GTC AAC ATC AGT CAC CCC GAT Gln Leu Arg Arg Gly Glu Arg Val Tyr Val Asn Ile Ser His Pro Asp 210 215 220	672
ATG GTG GAC TTC GCG AGA GGG AAG ACC TTC TTT GGG GGC GIG ATG GIG Met Val Asp Phe Ala Arg Gly Lys Thr Phe Phe Gly Ala Val Met Val 225 230 235 240	720
GGG TGA Gly	726

(2) INFORMATION FOR SEQ ID NO:2:

(1) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 241 amino acids
(B) TYPE: amino acid
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:

Leu Gly Leu Glu Gly Arg Gly Gly Arg Leu Gln Gly Arg Gly Ser Leu 1 5 10 15
Leu Leu Ala Val Ala Gly Ala Thr Ser Leu Val Thr Leu Leu Leu Ala 20 25 30
Val Pro Ile Thr Val Leu Ala Val Leu Ala Leu Val Pro Gln Asp Gln 35 40 45
Gly Gly Leu Val Thr Glu Thr Ala Asp Pro Gly Ala Gln Ala Gln Gln 50 55 60
Gly Leu Gly Phe Gln Lys Leu Pro Glu Glu Glu Pro Glu Thr Asp Leu 65 70 75 80
Ser Pro Gly Leu Pro Ala Ala His Leu Ile Gly Ala Pro Leu Lys Gly 85 90 95
Gln Gly Leu Gly Trp Glu Thr Thr Lys Glu Gln Ala Phe Leu Thr Ser 100 105 110
Gly Thr Gln Phe Ser Asp Ala Glu Gly Leu Ala Leu Pro Gln Asp Gly 115 120 125
Leu Tyr Tyr Leu Tyr Cys Leu Val Gly Tyr Arg Gly Arg Ala Pro Pro 130 135 140
Gly Gly Gly Asp Pro Gln Gly Arg Ser Val Thr Leu Arg Ser Ser Leu 145 150 155 160

SUBSTITUTE SHEET (RULE 26)

- 79 -

Tyr Arg Ala Gly Gly Ala Tyr Gly Pro Gly Thr Pro Glu Leu Leu Leu
 165 170 175
 Glu Gly Ala Glu Thr Val Thr Pro Val Leu Asp Pro Ala Arg Arg Gln
 180 185 190
 Gly Tyr Gly Pro Leu Trp Tyr Thr Ser Val Gly Phe Gly Gly Leu Val
 195 200 205
 Gln Leu Arg Arg Gly Glu Arg Val Tyr Val Asn Ile Ser His Pro Asp
 210 215 220
 Met Val Asp Phe Ala Arg Gly Lys Thr Phe Phe Gly Ala Val Met Val
 225 230 235 240
 Gly

(2) INFORMATION FOR SEQ ID NO:3:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 606 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: double
 (D) TOPOLOGY: linear

(ix) FEATURE:

- (A) NAME/KEY: CDS
 (B) LOCATION: 1..603

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:3:

CIG GOC TTA GIG OOC CAG GAT CAG GGA GGA CIG GTA ACG GAG ACG GGC Leu Ala Leu Val Pro Gln Asp Gln Gly Gly Leu Val Thr Glu Thr Ala 1 5 10 15	48
GAC OOC GGG GCA CAG GOC CAG CAA GGA CIG GGG TTT CAG AAG CIG CCA Asp Pro Gly Ala Gln Ala Gln Gln Gly Leu Gly Phe Gln Lys Leu Pro 20 25 30	96
GAG GAG GAG CCA GAA ACA GAT CTC AGC OOC GGG CTC CCA GGT GGC CAC Glu Glu Glu Pro Glu Thr Asp Leu Ser Pro Gly Leu Pro Ala Ala His 35 40 45	144
CTC ATA GGC GGT CCG CIG AAG GGG CAG GGG CTA GGC TGG GAG ACG ACG Leu Ile Gly Ala Pro Leu Lys Gly Gln Gly Leu Gly Trp Glu Thr Thr 50 55 60	192
AAG GAA CAG GCG TTT CIG ACG AGC GGG ACG CAG TTC TCG GAC GGC GAG Lys Glu Gln Ala Phe Leu Thr Ser Gly Thr Gln Phe Ser Asp Ala Glu 65 70 75 80	240
GGG CIG GCG CTC CCG CAG GAC GGC CTC TAT TAC CTC TAC TGT CTC GTC Gly Leu Ala Leu Pro Gln Asp Gly Leu Tyr Tyr Leu Tyr Cys Leu Val 85 90 95	288

SUBSTITUTE SHEET (RULE 26)

- 80 -

GGC TAC GCG GGC GCG GCG CCC OCT GGC GGC GCG GAC CCC CAG GGC GGC Gly Tyr Arg Gly Arg Ala Pro Pro Gly Gly Gly Asp Pro Gln Gly Arg 100 105 110	336
TGC GTC AGC CIG GGC AGC TCT CIG TAC GCG GCG GCG GGC GGC TAC GGC Ser Val Thr Leu Arg Ser Ser Leu Tyr Arg Ala Gly Gly Ala Tyr Gly 115 120 125	384
CCG GGC ACT CCC GAG CIG CIG CTC GAG GGC GGC GAG AGC GIG ACT CCA Pro Gly Thr Pro Glu Leu Leu Leu Glu Gly Ala Glu Thr Val Thr Pro 130 135 140	432
GIG CIG GAC CCG GGC AGG AGA CAA GCG TAC GCG OCT CTC TGG TAC AGC Val Leu Asp Pro Ala Arg Arg Gln Gly Tyr Gly Pro Leu Trp Tyr Thr 145 150 155 160	480
AGC GIG GCG TTC GGC GGC CIG GIG CAG CTC CCG AGG GGC GAG AGG GIG Ser Val Gly Phe Gly Gly Leu Val Gln Leu Arg Arg Gly Glu Arg Val 165 170 175	528
TAC GTC AAC ATC AGT CAC CCC GAT ATG GIG GAC TTC CCG AGA GCG AAG Tyr Val Asn Ile Ser His Pro Asp Met Val Asp Phe Ala Arg Gly Lys 180 185 190	576
ACC TTC TTT GCG GGC GIG ATG GIG GCG TGA Thr Phe Phe Gly Ala Val Met Val Gly 195 200	606

(2) INFORMATION FOR SEQ ID NO:4:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 201 amino acids
- (B) TYPE: amino acid
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:

Leu Ala Leu Val Pro Gln Asp Gln Gly Gly Leu Val Thr Glu Thr Ala 1 5 10 15
Asp Pro Gly Ala Gln Ala Gln Gln Gly Leu Gly Phe Gln Lys Leu Pro 20 25 30
Glu Glu Glu Pro Glu Thr Asp Leu Ser Pro Gly Leu Pro Ala Ala His 35 40 45
Leu Ile Gly Ala Pro Leu Lys Gly Gln Gly Leu Gly Trp Glu Thr Thr 50 55 60
Lys Glu Gln Ala Phe Leu Thr Ser Gly Thr Gln Phe Ser Asp Ala Glu 65 70 75 80

SUBSTITUTE SHEET (RULE 26)

- 81 -

Gly Leu Ala Leu Pro Gln Asp Gly Leu Tyr Tyr Leu Tyr Cys Leu Val
 85 90 95

Gly Tyr Arg Gly Arg Ala Pro Pro Gly Gly Gly Asp Pro Gln Gly Arg
 100 105 110

Ser Val Thr Leu Arg Ser Ser Leu Tyr Arg Ala Gly Gly Ala Tyr Gly
 115 120 125

Pro Gly Thr Pro Glu Leu Leu Leu Glu Gly Ala Glu Thr Val Thr Pro
 130 135 140

Val Leu Asp Pro Ala Arg Arg Gln Gly Tyr Gly Pro Leu Trp Tyr Thr
 145 150 155 160

Ser Val Gly Phe Gly Gly Leu Val Gln Leu Arg Arg Gly Glu Arg Val
 165 170 175

Tyr Val Asn Ile Ser His Pro Asp Met Val Asp Phe Ala Arg Gly Lys
 180 185 190

Thr Phe Phe Gly Ala Val Met Val Gly
 195 200

(2) INFORMATION FOR SEQ ID NO:5:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 450 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: double
- (D) TOPOLOGY: linear

(ix) FEATURE:

- (A) NAME/KEY: CDS
- (B) LOCATION: 1..447

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:5:

CCG CCG AAG GGG CAG GGG CTA GGC TGG GAG ACG ACG AAG GAA CAG GCG	48
Pro Leu Lys Gly Gln Gly Leu Gly Trp Glu Thr Thr Lys Glu Gln Ala	
1 5 10 15	
TTC CCG ACG AGC GGG ACG CAG TTC TCG GAC GGC GAG GGG CCG GCG CTC	96
Phe Leu Thr Ser Gly Thr Gln Phe Ser Asp Ala Glu Gly Leu Ala Leu	
20 25 30	
CCG CAG GAC GGC CTC TAT TAC CTC TAC TGT CTC GTC GGC TAC CCG GGC	144
Pro Gln Asp Gly Leu Tyr Tyr Leu Tyr Cys Leu Val Gly Tyr Arg Gly	
35 40 45	
CCG GCG CCC CCG GGC GGC GGC GAC CCC CAG GGC CCG TCG GTC ACG CTC	192
Arg Ala Pro Pro Gly Gly Gly Asp Pro Gln Gly Arg Ser Val Thr Leu	
50 55 60	

SUBSTITUTE SHEET (RULE 26)

- 82 -

GGC AGC TCT CTG TAC GGG GCG GGG GGC GGC TAC GGG CCG GGC ACT CCC Arg Ser Ser Leu Tyr Arg Ala Gly Gly Ala Tyr Gly Pro Gly Thr Pro 65 70 75 80	240
GAG CTG CTG CTC GAG GGC GGC GAG ACG GIG ACT CCA GIG CTG GAC CCG Glu Leu Leu Leu Glu Gly Ala Glu Thr Val Thr Pro Val Leu Asp Pro 85 90 95	288
GGC AGG AGA CAA GGG TAC GGG CCT CTC TGG TAC ACG AGC GIG GGG TTC Ala Arg Arg Gln Gly Tyr Gly Pro Leu Trp Tyr Thr Ser Val Gly Phe 100 105 110	336
GGC GGC CTG GIG CAG CTC CCG AGG GGC GAG AGG GIG TAC GTC AAC ATC Gly Gly Leu Val Gln Leu Arg Arg Gly Glu Arg Val Tyr Val Asn Ile 115 120 125	384
AGT CAC CCC GAT ATG GIG GAC TTC GCG AGA GGG AAG ACC TTC TTT GGG Ser His Pro Asp Met Val Asp Phe Ala Arg Gly Lys Thr Phe Phe Gly 130 135 140	432
GGC GIG ATG GIG GGG TGA Ala Val Met Val Gly 145	450

(2) INFORMATION FOR SEQ ID NO:6:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 149 amino acids
- (B) TYPE: amino acid
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:6:

Pro Leu Lys Gly Gln Gly Leu Gly Trp Glu Thr Thr Lys Glu Gln Ala 1 5 10 15
Phe Leu Thr Ser Gly Thr Gln Phe Ser Asp Ala Glu Gly Leu Ala Leu 20 25 30
Pro Gln Asp Gly Leu Tyr Tyr Leu Tyr Cys Leu Val Gly Tyr Arg Gly 35 40 45
Arg Ala Pro Pro Gly Gly Gly Asp Pro Gln Gly Arg Ser Val Thr Leu 50 55 60
Arg Ser Ser Leu Tyr Arg Ala Gly Gly Ala Tyr Gly Pro Gly Thr Pro 65 70 75 80
Glu Leu Leu Leu Glu Gly Ala Glu Thr Val Thr Pro Val Leu Asp Pro 85 90 95
Ala Arg Arg Gln Gly Tyr Gly Pro Leu Trp Tyr Thr Ser Val Gly Phe 100 105 110

SUBSTITUTE SHEET (RULE 26)

- 83 -

Gly Gly Leu Val Gln Leu Arg Arg Gly Glu Arg Val Tyr Val Asn Ile
 115 120 125

Ser His Pro Asp Met Val Asp Phe Ala Arg Gly Lys Thr Phe Phe Gly
 130 135 140

Ala Val Met Val Gly
 145

(2) INFORMATION FOR SEQ ID NO:7:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 156 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: double
 (D) TOPOLOGY: linear

(ix) FEATURE:

- (A) NAME/KEY: CDS
 (B) LOCATION: 1..156

(x) SEQUENCE DESCRIPTION: SEQ ID NO:7:

CIG GGC TTA GIG CCC CAG GAT CAG GGA GGA CIG GTA ACG GAG ACG GGC	48
Leu Ala Leu Val Pro Gln Asp Gln Gly Gly Leu Val Thr Glu Thr Ala	
1 5 10 15	
GAC CCC GGG GCA CAG GGC CAG CAA GGA CIG GGG TTT CAG AAG CIG CCA	96
Asp Pro Gly Ala Gln Ala Gln Gln Gly Leu Gly Phe Gln Lys Leu Pro	
20 25 30	
GAG GAG GAG CCA GAA ACA GAT CTC AGC CCC GGG CTC CCA GCT GGC CAC	144
Glu Glu Glu Pro Glu Thr Asp Leu Ser Pro Gly Leu Pro Ala Ala His	
35 40 45	
CTC ATA GGC GGT	
Leu Ile Gly Ala	156
50	

(2) INFORMATION FOR SEQ ID NO:8:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 52 amino acids
 (B) TYPE: amino acid
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(x) SEQUENCE DESCRIPTION: SEQ ID NO:8:

Leu Ala Leu Val Pro Gln Asp Gln Gly Gly Leu Val Thr Glu Thr Ala
1 5 10 15

SUBSTITUTE SHEET (RULE 26)

- 84 -

Asp Pro Gly Ala Gln Ala Gln Gln Gly Leu Gly Phe Gln Lys Leu Pro
 20 25 30
 Glu Glu Glu Pro Glu Thr Asp Leu Ser Pro Gly Leu Pro Ala Ala His
 35 40 45
 Leu Ile Gly Ala
 50

(2) INFORMATION FOR SEQ ID NO:9:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 17 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(iii) HYPOTHEICAL: NO

(iv) ANTI-SENSE: YES

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:9:

GTTTGGGCT CTTCTTC

17

(2) INFORMATION FOR SEQ ID NO:10:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 17 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(iii) HYPOTHEICAL: NO

(iv) ANTI-SENSE: YES

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:10:

GTTTGGGCT CTTCTTC

17

(2) INFORMATION FOR SEQ ID NO:11:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 18 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

SUBSTITUTE SHEET (RULE 26)

- 85 -

(ii) MOLECULE TYPE: cDNA

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: NO

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:11:

ATGGGGGGCAC TGGGGCTG

18

(2) INFORMATION FOR SEQ ID NO:12:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 19 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: double

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: NO

(ix) FEATURE:

(A) NAME/KEY: misc feature

(B) LOCATION: 1..19

(D) OTHER INFORMATION: /function= "linker"

(ix) FEATURE:

(A) NAME/KEY: misc feature

(B) LOCATION: 1..19

(D) OTHER INFORMATION: /note= "The 5' terminal 4 nucleotides in the complementary strand are not present in this linker "

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:12:

GCGGGGGCTT TAGAGCACA

19

(2) INFORMATION FOR SEQ ID NO:13:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 27 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: NO

SUBSTITUTE SHEET (RULE 26)

- 86 -

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:13:

GCACGTCGATA GGCACCGGCGCA GCACAA

27

SUBSTITUTE SHEET (RULE 26)

86/1

Applicant's or agent's file reference number	B129 CIP II	International application No.
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INDICATIONS RELATING TO A DEPOSITED MICROORGANISM

(PCT Rule 13bis)

A. The indications made below relate to the microorganism referred to in the description on page 29, line at footnote, page 51, lines 9-23, and page 72, line 5 through page 73, line 25	
B. IDENTIFICATION OF DEPOSIT	
Further deposits are identified on an additional sheet <input type="checkbox"/> E. coli K12 bearing plasmid with insert coding Name of depository institution for protein P33 (LT-beta), BN1289(MC1061/P3/ P33-clone 12) American Type Culture Collection Address of depository institution (including postal code and country) 12301 Parklawn Drive Rockville, Maryland 20852 United States of America	
Date of deposit 16 November 1992 (16.11.92)	Accession Number 69125
C. ADDITIONAL INDICATIONS (leave blank if not applicable) This information is contained on an additional sheet <input type="checkbox"/>	
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- 87 -

Claims

We claim:

1. Lymphotoxin- β , a lymphocyte membrane-type polypeptide comprising SEQ ID NO:2.
2. The polypeptide according to claim 1 wherein the polypeptide is associated with a cell surface.
3. The polypeptide according to claim 2 wherein the polypeptide is associated with the surface of OKT3-stimulated primary T cells, antigen-specific IL-2 dependent CTL clones, and a PMA-stimulated human T cell hybridoma, II-23.D7.
4. A soluble lymphotoxin- β peptide comprising an amino acid sequence selected from the group consisting of:
 - (a) SEQ ID NO:4;
 - (b) SEQ ID NO:6; and
 - (c) an amino acid sequence represented by the following formula:
$$X - \text{SEQ ID NO:6},$$
wherein X comprises one or more of the amino acid residues starting from the 3' end of SEQ ID NO:8.
5. A peptide according to claim 4 further comprising a leader sequence at the 5' end.
6. A polypeptide comprising an amino acid sequence that is encoded by a DNA from the group consisting of:
 - (a) a DNA sequence comprising SEQ ID NO:1;

SUBSTITUTE SHEET (RULE 26)

- 88 -

(b) DNA sequences that hybridize to the DNA defined by SEQ ID NO:1 and that code on expression for a polypeptide that is substantially homologous with lymphotoxin-8; and

(c) DNA comprising degenerate nucleotide sequences that code for the polypeptide that is encoded by the DNA sequence defined by SEQ ID NO:1.

7. A polypeptide comprising an amino acid sequence that is encoded by a DNA from the group consisting of:

(a) a DNA sequence comprising SEQ ID NO:3;

(b) a DNA sequence comprising SEQ ID NO:5;

(c) a DNA sequence represented by the following formula:

X - SEQ ID NO:5,

wherein X comprises one or more of the nucleoside triplets starting from the 3' end of SEQ ID NO:7;

(d) DNA sequences that hybridize to any one of SEQ ID NO:3, SEQ ID NO:5 and the sequence according to part (c) above and that code on expression for a polypeptide that is substantially homologous with a soluble lymphotoxin-8 peptide; and

(e) a DNA sequence comprising degenerate nucleotide sequences that code for the polypeptide encoded for any one of SEQ ID NO:3, SEQ ID NO:5 and the sequence according to part (c) above.

8. An engineered polypeptide comprising the amino acid sequence defined by SEQ ID NO:2 wherein the sequence Leu Gly Leu is cleaved from the 5' end of said sequence and replaced by a single Met or Leu residue.

SUBSTITUTE SHEET (RULE 26)

- 89 -

9. An isolated DNA sequence selected from the group consisting of:

- (a) a DNA sequence comprising the nucleotide sequence defined by SEQ ID NO:1;
- (b) a DNA sequence that hybridizes with the DNA sequence defined by SEQ ID NO:1 and that codes on expression for a polypeptide that is substantially homologous with lymphotoxin- β ; and
- (c) a DNA sequence comprising degenerate nucleotide sequences that code for lymphotoxin- β .

10. An isolated DNA sequence selected from the group consisting of:

- (a) a DNA sequence comprising the nucleotide sequence defined by SEQ ID NO:3;
- (b) a DNA sequence comprising the nucleotide sequence defined by SEQ ID NO:5;
- (c) a DNA sequence comprising the nucleotide sequence according to claim 7(c);
- (d) DNA sequences that hybridize to a DNA sequence as defined by any one of SEQ ID NO:3, SEQ ID NO:5 or the sequence according to claim 7(c) and that code on expression for a polypeptide that is substantially homologous with a soluble lymphotoxin- β peptide; and
- (e) a DNA sequence comprising degenerate nucleotide sequences that code for a soluble lymphotoxin- β peptide.

11. An engineered DNA sequence comprising the nucleotide sequence defined by SEQ ID NO:1 wherein the nucleotides CTGGGCTG are cleaved from the 5' end of said sequence and replaced by a single start codon.

SUBSTITUTE SHEET (RULE 26)

- 90 -

12. A recombinant DNA molecule comprising a DNA sequence selected from the group consisting of:

- (a) a DNA sequence defined by SEQ ID NO:1;
- (b) a DNA sequence defined by SEQ ID NO:3;
- (c) a DNA sequence defined by SEQ ID NO:5;
- (d) a DNA sequence according to claim 7(c);
- (e) a DNA sequence according to Claim 11;
- (f) a DNA sequence that hybridizes with the DNA sequences defined by any one of SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:5 and the sequence according to claim 7(c) and that codes on expression for lymphotoxin- β or a soluble lymphotoxin- β peptide;
- (g) a DNA sequence comprising degenerate nucleotide sequences that codes for lymphotoxin- β ; and
- (h) a DNA sequence comprising degenerate nucleotide sequences that codes for a soluble lymphotoxin- β peptide.

13. A host selected from the group consisting of unicellular hosts, animal cells in culture and human cells in culture, transfected with the recombinant DNA molecule of claim 12.

14. The host according to claim 13 selected from the group of tumor infiltrating lymphocytes, lymphokine activated killer cells, killer cells and genetically engineered tumor cells removed from a patient.

SUBSTITUTE SHEET (RULE 26)

- 91 -

15. A method for producing the polypeptide of any one of claims 1 to 8, said method comprising the steps of culturing a transformed host according to claim 13 and collecting the polypeptide.

16. A polypeptide complex comprising a first polypeptide selected from a group consisting of the amino acid sequence defined by any one of SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:6, a polypeptide according to claim 8, and soluble lymphotoxin- β peptide according to claim 4(c), and a second polypeptide selected from the group consisting of lymphotoxin- α , native human or animal lymphotoxin, recombinant lymphotoxin, soluble lymphotoxin, secreted lymphotoxin, or lymphotoxin or lymphotoxin-active fragments of any of the above.

17. A polypeptide complex comprising a plurality of lymphotoxin- β polypeptide units.

18. A polypeptide complex according to claim 16 wherein the complex is associated with a cell surface.

19. A polypeptide complex according to claim 18 wherein the first polypeptide is associated with the surface of OKT3-stimulated primary T cells, antigen-specific IL-2 dependent CTL clones, and a PMA-stimulated non-lymphotoxin human T cell hybridoma, II-23.D7.

20. A method for producing lymphotoxin epitopes on the surface of a cell comprising the steps of transfecting the cell with a recombinant DNA molecule according to claim 12 and expressing that DNA in the cell.

SUBSTITUTE SHEET (RULE 26)

- 92 -

21. A method for enhancing the targeting tumoricidal activity of tumor infiltrating lymphocytes comprising the steps of transfecting the lymphocytes with a recombinant DNA molecule according to claim 12 and introducing the transformed lymphocytes to a patient.

22. The method according to claim 21, wherein the transformed lymphocytes are incubated with a lymphokine before or after transfection with the recombinant DNA molecule according to claim 12.

23. The method according to claim 22, wherein the lymphokine is IL-2.

24. A composition for preventing, treating or lessening the advancement, severity or effects of HIV infection, neoplasia, inflammation or inflammatory disease, or autoimmune disease comprising an effective amount of a polypeptide selected from the group consisting of a polypeptide according to any one of claims 1 to 8, a polypeptide complex according to any one of claims 16-19, antibodies to any one of the above, or a combination of any of the above, and a pharmaceutically acceptable carrier.

25. A method for preventing, treating or lessening the advancement, severity or effects of HIV infection, neoplasia, inflammation or inflammatory diseases, or autoimmune disease comprising administering an effective amount of a polypeptide selected from the group consisting of a polypeptide according to any one of claims 1-8, a polypeptide complex according to any one of claims 16-19,

SUBSTITUTE SHEET (RULE 26)

- 93 -

antibodies to any one of the above, or a combination of any of the above, and a pharmaceutically acceptable carrier.

26. A composition for suppressing the immune system comprising an effective amount of a polypeptide selected from the group consisting of a polypeptide according to any one of claims 1 to 8, a polypeptide complex according to any one of claims 16-19, antibodies to any one of the above, or a combination of any of the above, and a pharmaceutically acceptable carrier.

27. A method for suppressing the immune system comprising administering an effective amount of a polypeptide selected from the group consisting of a polypeptide according to any one of claims 1-8, a polypeptide complex according to any one of claims 16-19, antibodies to any one of the above, or a combination of any of the above, and a pharmaceutically acceptable carrier.

28. A nucleotide sequence coding for lymphotoxin-8 comprising the nucleotide sequence represented by SEQ ID NO:1 and further comprising an engineered nucleotide sequence at the 5' end wherein said engineered sequence comprises a functional start codon that is either ATG or CTG and wherein any other codon within said engineered sequence coding for leucine is not CTG.

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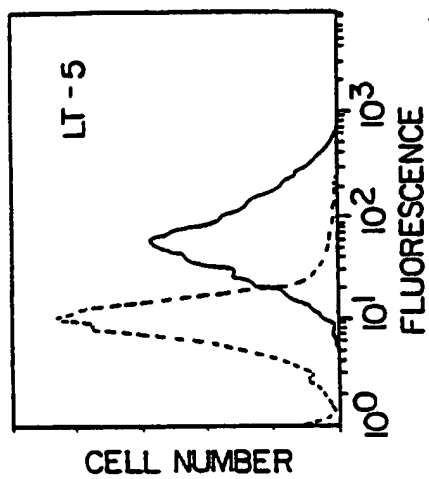


FIG. 1B

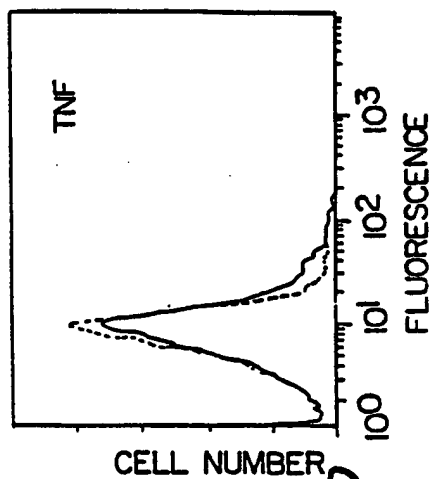


FIG. 1D

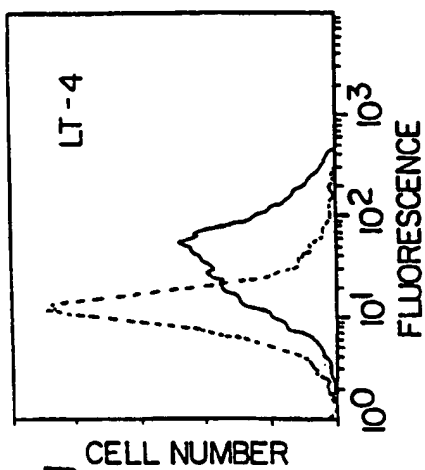


FIG. 1A

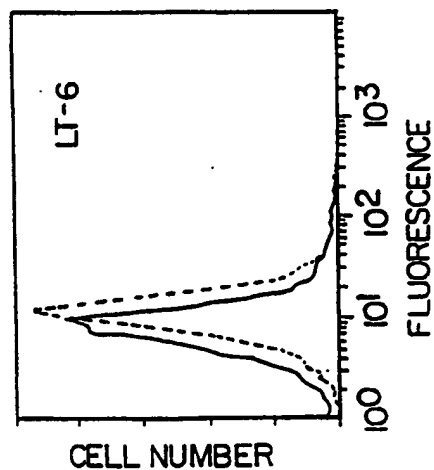


FIG. 1C

2/16

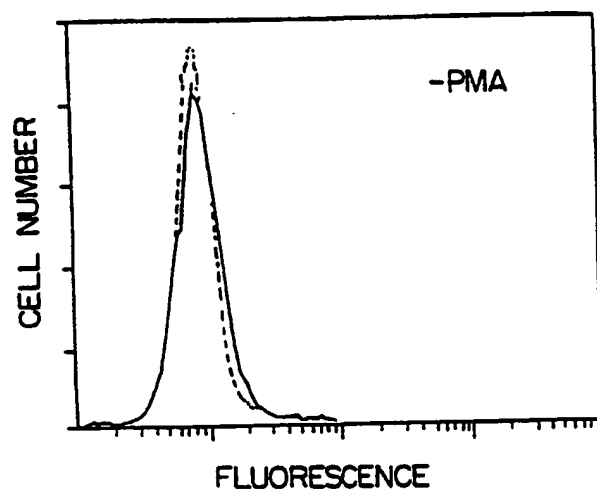


FIG. 2A

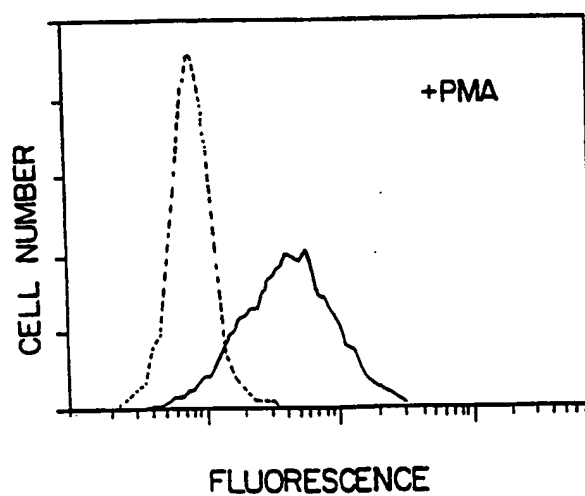


FIG. 2B

3/16

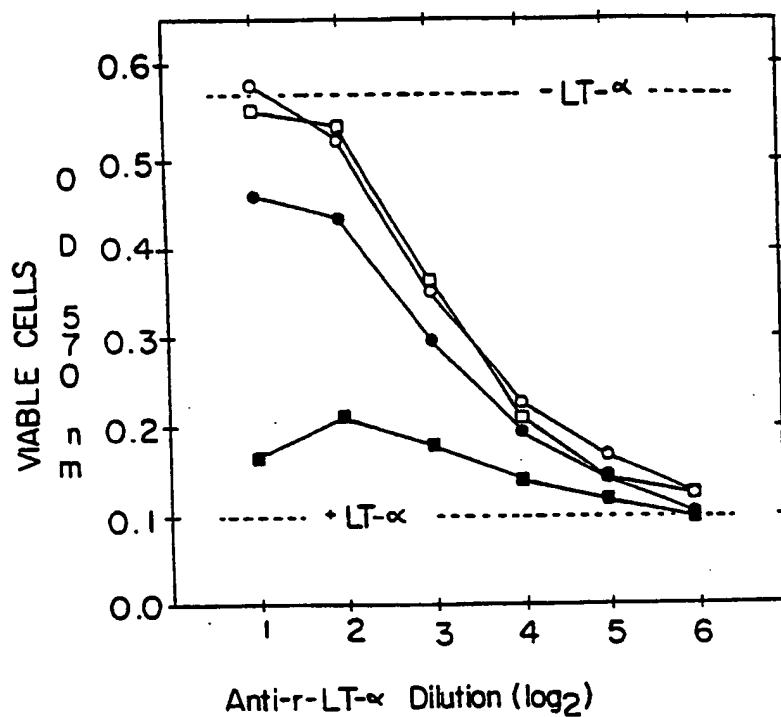


FIG. 3

4/16

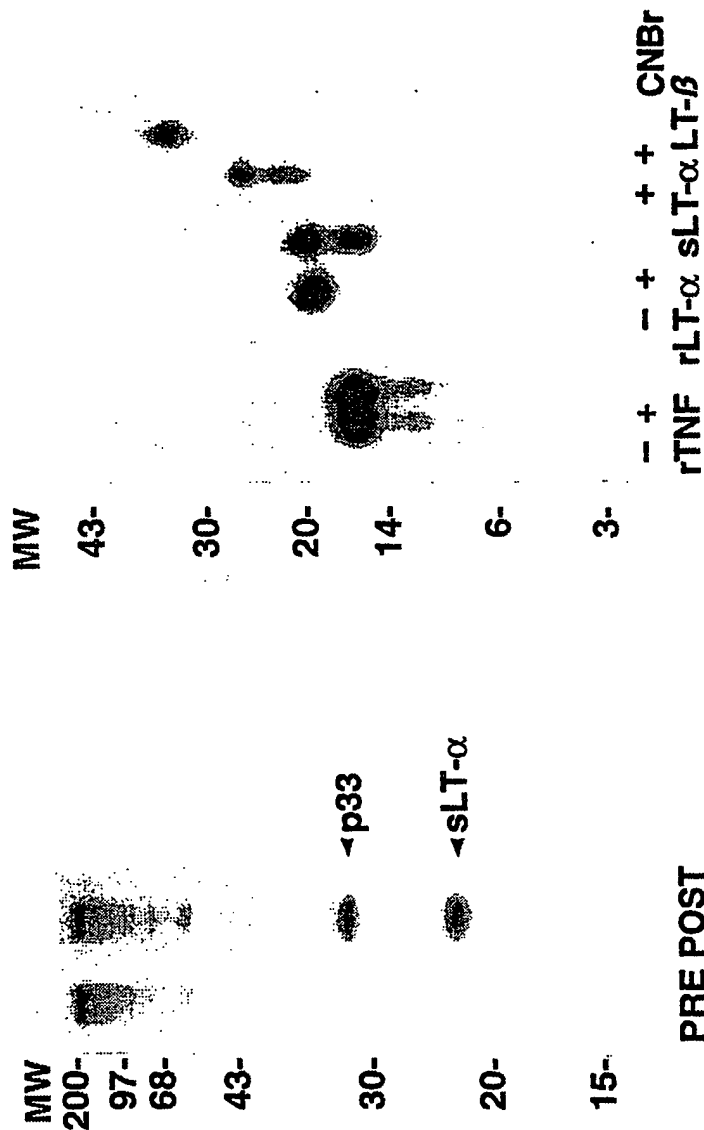


Fig. 4B

Fig. 4A

5/16

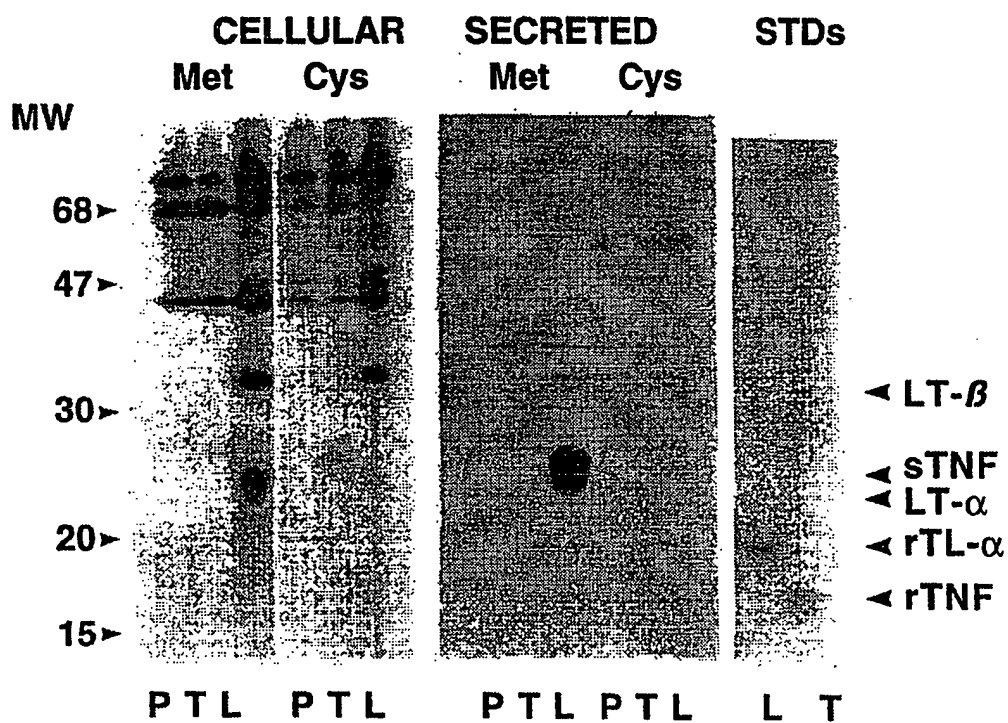


FIG. 5

6/16

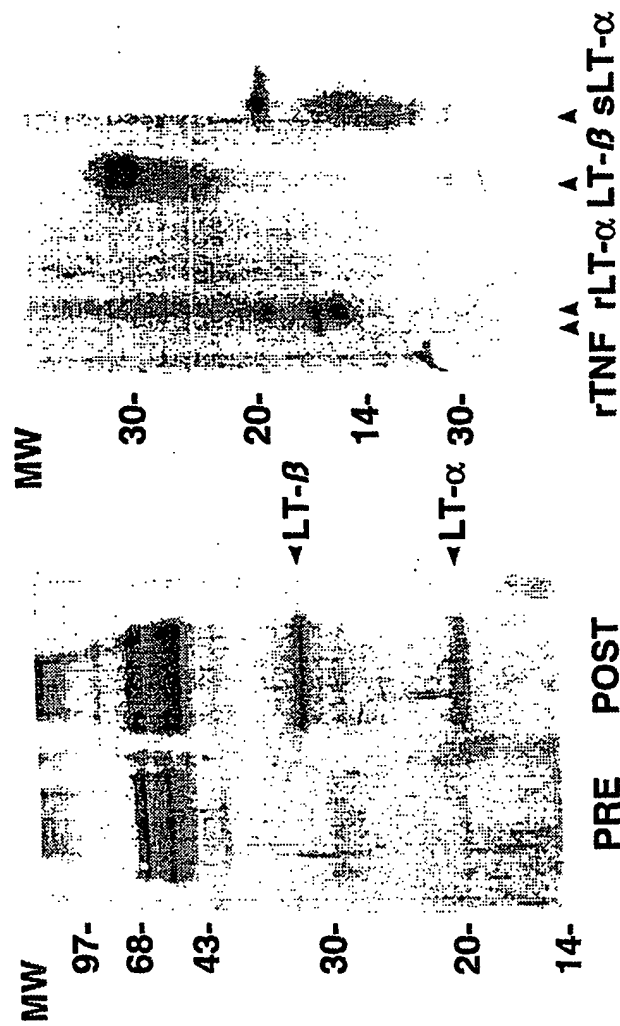
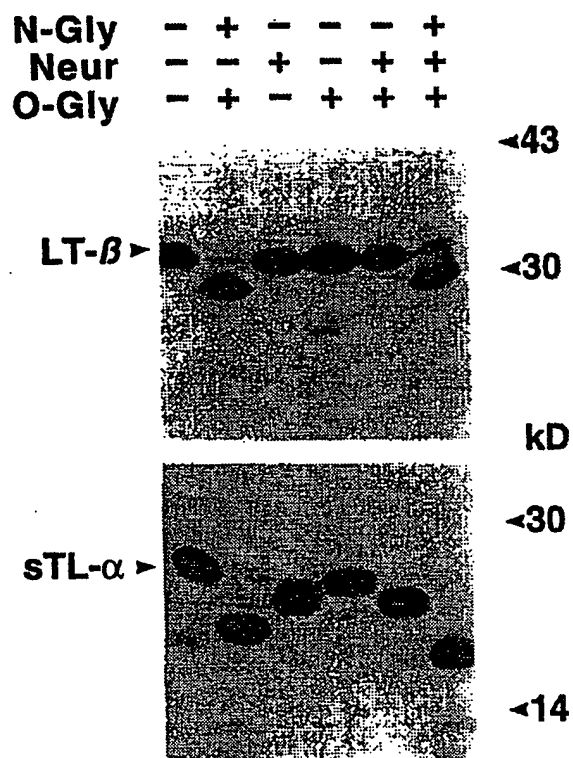


FIG. 6B

FIG. 6A

7 / 16

**Fig. 7**

8/16

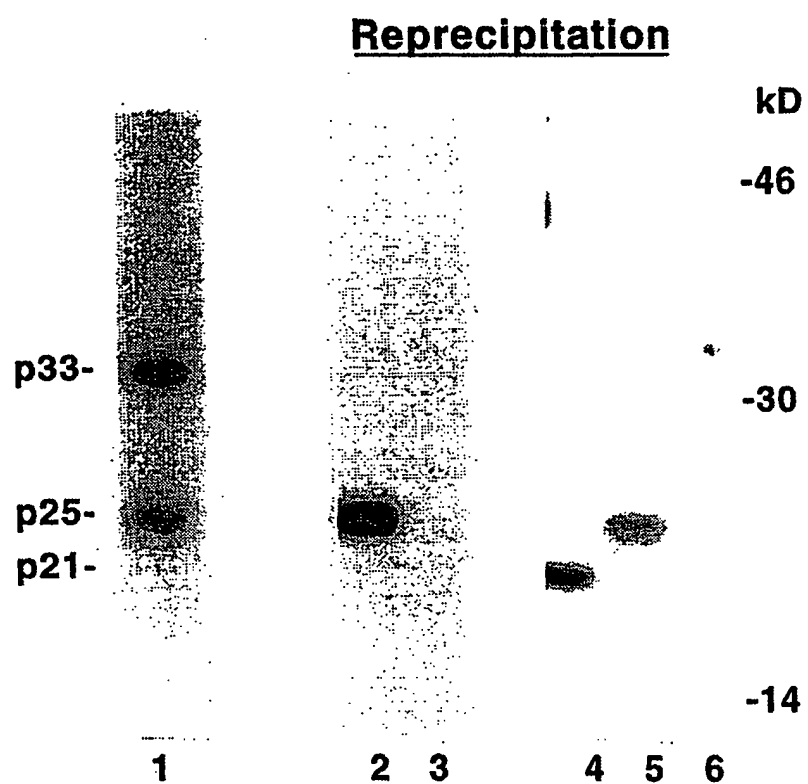


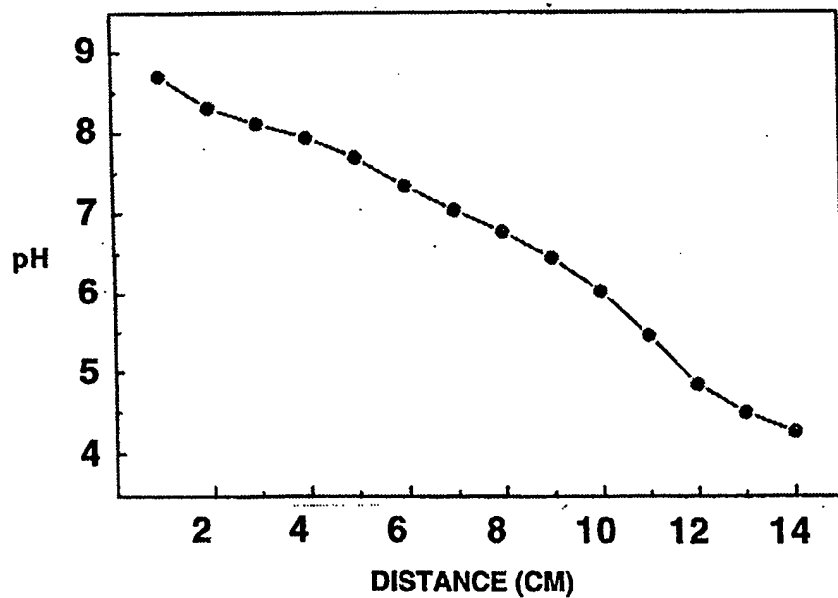
Fig. 8

9/16

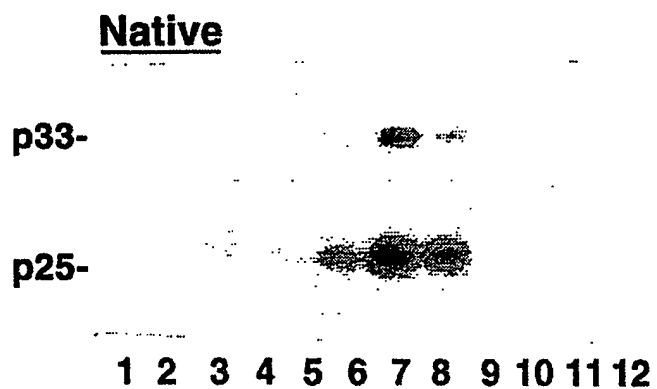
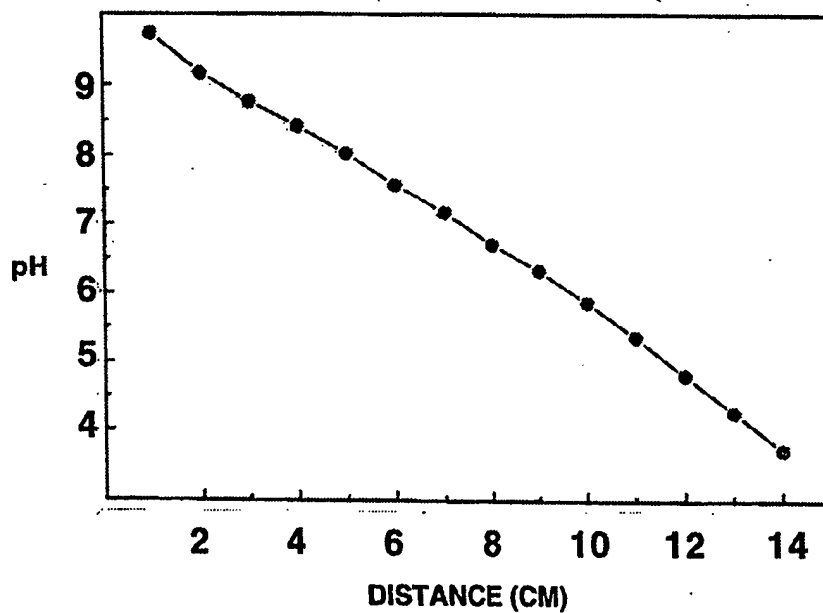
Denatured

p33-

p25-

Fig. 9A***Fig. 9B***

10/16

**Fig. 10A****Fig. 10B**

11/16

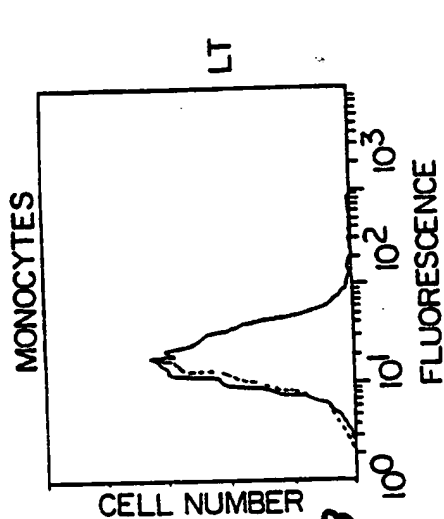


FIG. 11B

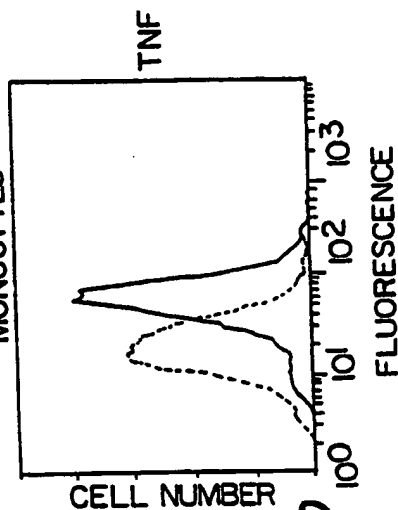


FIG. 11D

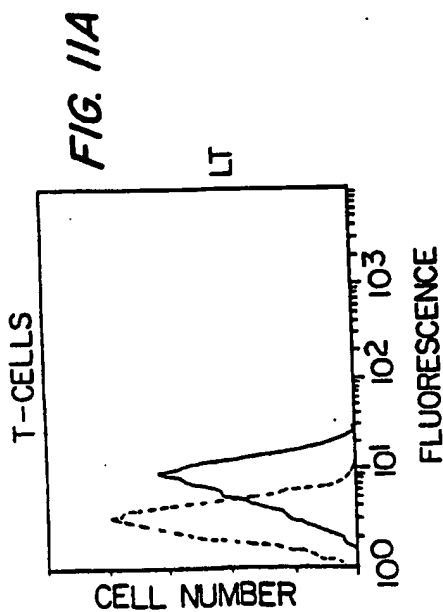


FIG. 11A

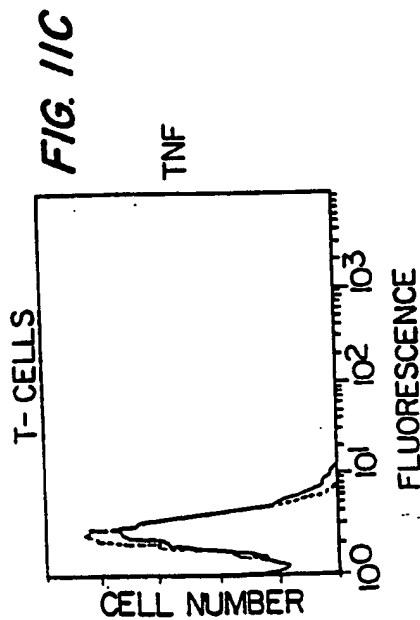


FIG. 11C

12/16

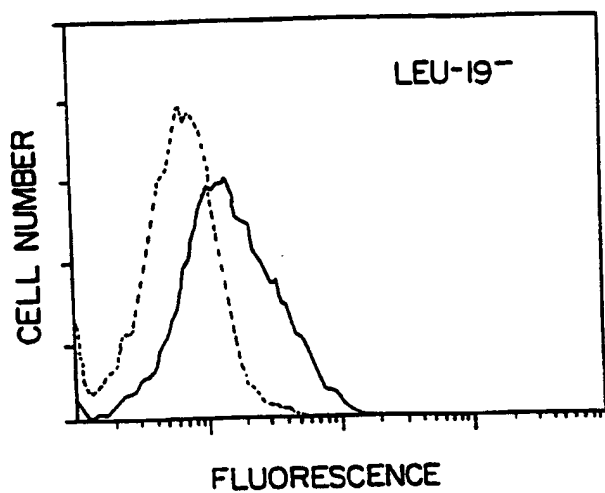


FIG. 12A

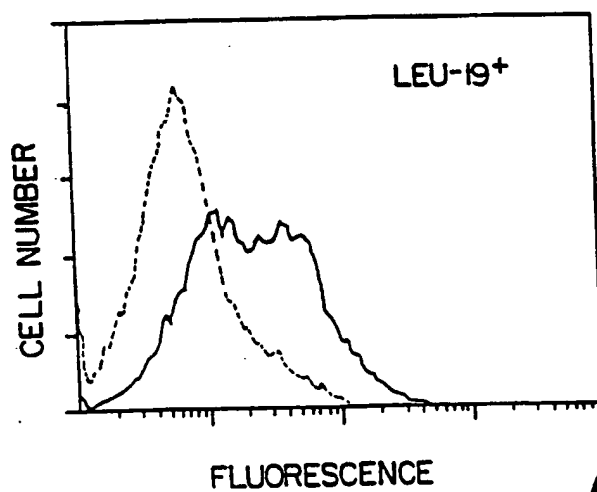


FIG. 12B

13/16

LT- β Amino Acid Sequences

N-terminus:

GLEGRGxRLQGRGSLLAVAGATGLVTLL--VPI

Tryptic Peptides

T87/88:

LP E E E P E T D L S P G L P A A H L I G A P L K

T110:

x Q A F L T S G T Q F S D A E G L A L P Q D G L Y

T67:

X Q G L x x E T

T48:

S S L Y R

T96:

A G G A Y G P G T P E L L L L E G A E T V T P V L D

Fig. 13

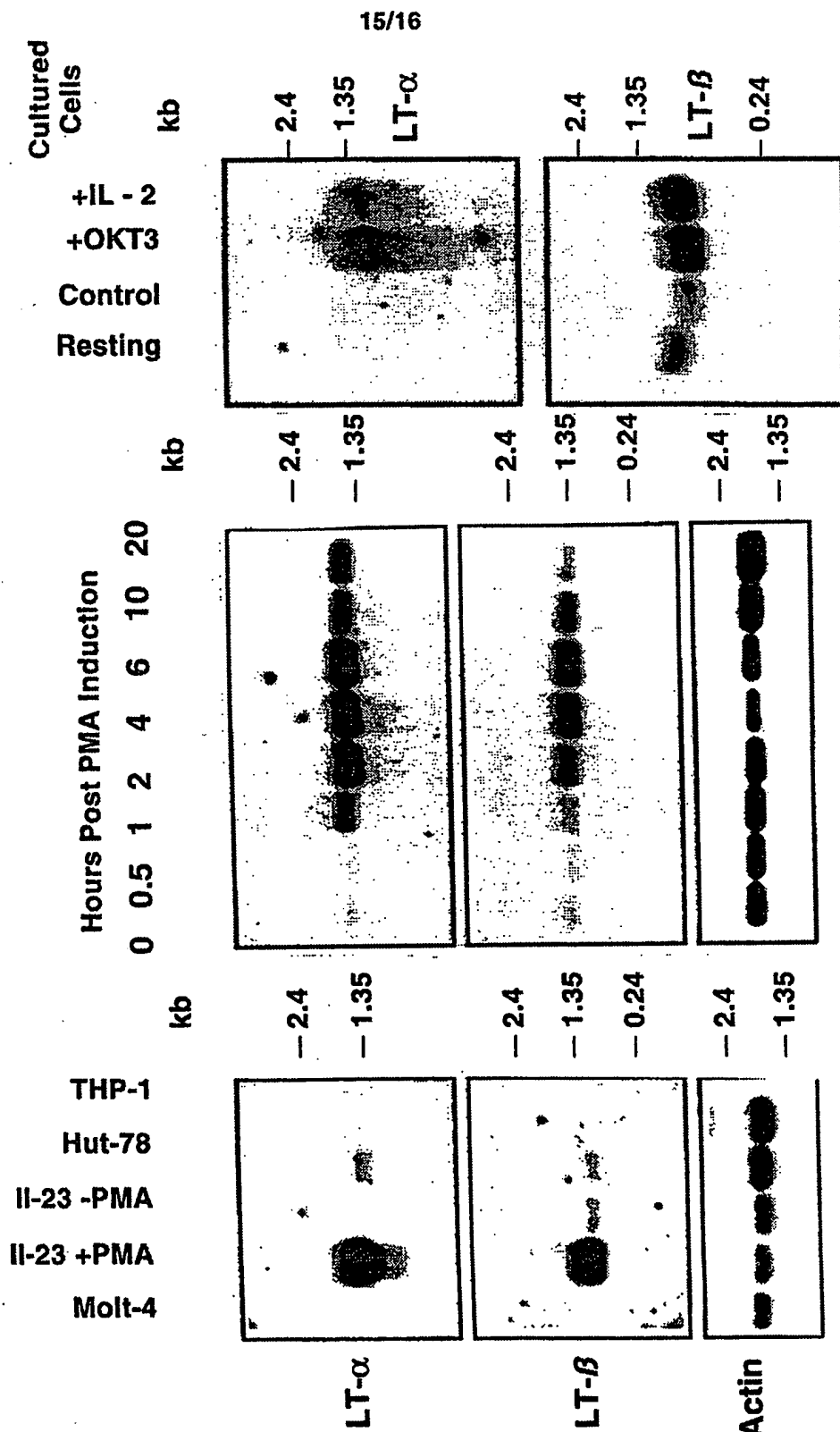
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14/16

SEQUENCE COMPARISON OF VARIOUS LIGANDS IN THE TNF-R FAMILY

	1	10	20	30	50	60
hTNF						
hTNF	MSTESMIRD					
mCD40L	VELAEEALPKKTGGPQGSRRCLFSLSFSLIVAGATTLECLLHFGVIGPQ					
hP33	MIETYSQSPRSVATGLPASMKIFMYLLTVFLITOMIGSVLFAVYLHRR					
hLT	GLEGRGGRLQGRGSLLLAVAGATSLVTLLLAPEITVLAVLALVPQDQ					
	MTPPERLFLPRVCGTTLHLLLLGLLLVLLPGAQGLPGVGLT					
hTNF	-----REE-FPRD-----					
mCD40L	LDKVEEEVNLHEDFVF IKKLKRCNKGEGLSLNCEEMRRQFEDLVKDIT					
hP33	GGLVTETADPGAQAQ-----					
hLT	-----					
hTNF	LSLISPLAQAVRSSSRTPSDKPVAVHVVANPQAEQ--LQWLNRRANALLA					
mCD40L	LNKEEKKENSFEMQRGDEDPQIAAHVVSEANSNAASVLQWAKKGYITMKS					
hP33	QGLGFQKLP EEEFETDLSPLGLPAHLLIGAPLK-GQ-GLGWETTKEQAFLT					
hLT	PSAAQTARQHFKMHLAHS TLKPAHLLIGDPSK--QNSLLWRANTDRAFLQ					
hTNF	NGVELRD-NQLVVPSEGLYLIYSQVLFKGQGCPSTHVLLTHTISRIAVSY					
mCD40L	NLVMLENGKQLTVKREGLYYVYTQVTFCSNREPSSQRPFI VGLWLKPSIG					
hP33	SGTQFSDAEGALPQDGLYYLYCLVGYRGRAPPGGDPQGRSVTLRSSLY					
hLT	DGFSLSNNS-LLVPTSGIYFVYSQVVFSGKAYS PKATSSPLYLAHEVQLF					
hTNF	QTKVN-----LLSAIKSPCQRETPEGAEAK--P-WYEP IYLGGVFQLE					
mCD40L	SER-----ILLKAANTHSSSQLCEQQ-----SVHLGGVFELQ					
hP33	RAGGAYGPGTPELLEGAETVTPVLD PARRQGYGPLWYTSVGF GGLVQLR					
hLT	SS--QYPFHVP-LLSSQK-MVYPGLQE-----P-WLHSMYHGAATQLT					
hTNF	KGDRLSAEINRPDY-LDFAESGQVYFGIIL					
mCD40L	AGASVFVNVT--EASQVIHRVGFSSFGLLKL					
hP33	RGERVYVNI SHPD-MVDFAR-GKTFFGAVMVG					
hLT	QGDQLSTHT--DGIPHLVLSPTVFFGAFAL					

Fig. 14

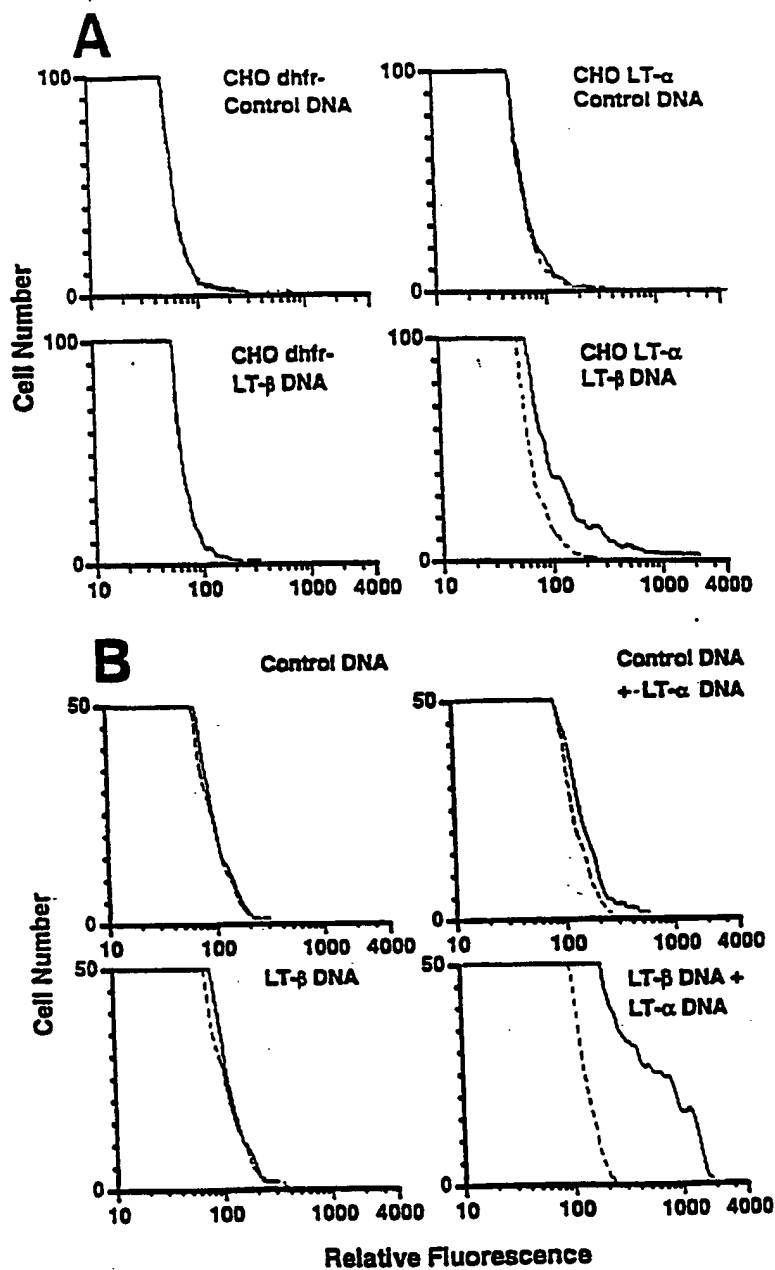


C.

B.
Fig. 15

A.

16/16

**Fig. 16**